Structure, Function, and Phylogeny of the Mating Locus in the *Rhizopus oryzae* Complex

Andrii P. Gryganskyi1,2,*, Soo Chan Lee3, Anastasia P. Litvintseva3, Matthew E. Smith1, Gregory Bonito1, Teresita M. Porter1, Iryna M. Anishchenko2, Joseph Heitman3, Rytas Vilgalys1

1 Department of Biology, Duke University, Durham, North Carolina, United States of America, 2 Department of Mycology, M.G. Kholodny Institute of Botany, National Academy of Sciences of Ukraine, Kyiv, Ukraine, 3 Department of Molecular Genetics and Microbiology, Duke University Medical Center, Durham, North Carolina, United States of America

### Abstract

The *Rhizopus oryzae* species complex is a group of zygomycete fungi that are common, cosmopolitan saprotrophs. Some strains are used beneficially for production of Asian fermented foods but they can also act as opportunistic human pathogens. Although *R. oryzae* reportedly has a heterothallic (+/−) mating system, most strains have not been observed to undergo sexual reproduction and the genetic structure of its mating locus has not been characterized. Here we report on the mating behavior and genetic structure of the mating locus for 54 isolates of the *R. oryzae* complex. All 54 strains have a mating locus similar in overall organization to *Phycomyces blakesleeanus* and *Mucor circinelloides* (Mucoromycotina, Zygomycota). In all of these fungi, the minus (−) allele features the SexM high mobility group (HMG) gene flanked by a RNA helicase gene and a TP transporter gene (TPT). Within the *R. oryzae* complex, the plus (+) mating allele includes an inserted region that codes for a BTB/POZ domain gene and the SexP HMG gene. Phylogenetic analyses of multiple genes, including the mating loci (HMG, TPT, RNA helicase), ITS1-5.8S-ITS2 rDNA, RPB2, and LDH genes, identified two distinct groups of strains. These correspond to previously described sibling species *R. oryzae* sensu stricto and *R. delemar*. Within each species, discordant gene phylogenies among multiple loci suggest an outcrossing population structure. The hypothesis of random-mating is also supported by a 50:50 ratio of plus and minus mating types in both cryptic species. When crossed with tester strains of the opposite mating type, most isolates of *R. delemar* failed to produce zygospores, while isolates of *R. oryzae* produced sterile zygospores. In spite of the reluctance of most strains to mate in vitro, the conserved sex locus structure and evidence for outcrossing suggest that a normal sexual cycle occurs in both species.

---

**Introduction**

*Rhizopus oryzae* is a complex of closely related, heterothallic species [1,2,3,4] that are common, cosmopolitan saprotrophs in soil, dung, and rotting vegetation [5,6,7,8]. Strains of the *R. oryzae* complex have been used for centuries as fermented food starters for the production of tempeh and other Asian foods [24,25,26]. In addition, some species produce azygos- pores, which are thought to be asexual. They are morphologically similar to zygospores but develop in the absence of a mating partner. Recent studies of sexual reproduction in the zygomycete *Phycomyces blakesleeanus* revealed that the mating system of this species is regulated by divergent alleles of a single gene: SexH (−) and SexP (+) [16]. The sex gene is a member of the high mobility genes (HMG) family and is located between two flanking genes that code for a triose-phosphate transporter homolog (TPT) and an RNA helicase [16]. A similar sex locus structure was also reported in another zygomycete, *Mucor circinelloides* [27] and was predicted for *R. oryzae* based on BLAST analysis of the publicly available genome of strain RA99-880 [16]. A recent study also found a similar sex-related locus in three Microsorid species, as the “*Rhizopus* pattern”. In this type of zygospore formation, some nuclei degenerate and meiosis is delayed until zygospore germination [21,23]. Zygospores have been observed in both heterothallic and homothallic *Rhizopus* species [3,4] but in most cases they do not germinate. Although zygospore germination is rare, it has been documented for *R. stolonifer* [24,25,26]. In addition, some *Rhizopus* species produce azygos-pores, which are thought to be asexual. The authors have declared that no competing interests exist.

* E-mail: apg10@duke.edu

---


**Editor:** Darren P. Martin, Institute of Infectious Disease and Molecular Medicine, South Africa

**Received July 28, 2010; Accepted November 4, 2010; Published December 9, 2010**

Copyright: © 2010 Gryganskyi et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This work was supported in part by National Institutes of Health (NIH)/National Institute of Allergy and Infectious Diseases R01 grant AI50113 to J. H. and R21 grant AI085331 to J. H. S. C. L. was supported by the NIH Molecular Mycology and Pathogenesis Training Program (AI52080). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.
which are believed to be closely related to the zygomycetes [27,28].

In this study, we characterized the sex locus in six strains of the R. oryzae species complex (four (+) strains and two (+) strains) and examined the genetic structure of the sex locus in a larger collection of clinical, industrial, and environmental isolates. We identified the most conserved regions in the sex locus and designed primers for three key regions: the high mobility group (HMG) gene regions, the triose-phosphate transporter (TPT) region, and the RNA helicase region. We used these primers to amplify fragments of the sex locus in 40 additional isolates and used these DNA sequences to study phylogenetic relationships among loci. Here we compare the phylogenetic pattern from the sex genes with those from several other commonly sequenced DNA loci (ITS1-5.8S-ITS2 and 28S rDNA, RPB2, mtSSU) and demonstrate that the sex loci are informative for differentiating the cryptic sibling species R. oryzae s. s. and R. delemar. We also compare the sex locus structure of R. oryzae (+) and (−) strains with those of the two mucoralean fungi Phymyogyes blakesleeanus and Mucor circinelloides and show that the sex locus of all three taxa has a similar overall arrangement.

Results

Mating tests produce sexual spores but not progeny

We repeated the mating tests reported by Schipper [3] using the same strains: (+) CBS346.36 and (−) CBS110.17, CBS112.07 (holotype culture), CBS127.08, CBS148.22, CBS257.28, CBS264.28, CBS266.30, CBS292.47, and CBS382.52. We were not able to include the holotype culture of Rhizopus delemar (CBS120.12, GenBank # AB181318), but we have included the R. delemar strain NRRRL1447, which has an ITS1-5.8S-ITS2 sequence that is identical to that of the type culture (Fig. S1A). As observed by Schipper, zygospores formed in all mating reactions except those performed with (−) strain CBS257.28.

We also tested additional R. oryzae isolates from the NRRL and Duke collections using the tester strains CBS346.36 (+) and CBS112.07 (−). These mating tests identified six additional compatible isolates that were capable of producing zygospores when paired with tester strains of the opposite mating type (Table 1, Table S3). Self-pairings or pairings with senescent cultures consistently failed to produce zygospores. Mating test results were not influenced by medium type, medium nutrient concentration, or Petri plate size. Most strains did not produce zygospores with either “plus” or “minus” testers under the conditions we tested. The genome strain RA99-880 also failed to produce zygospores in all mating tests with any of the 55 strains used in this study.

Where mating occurred, zygospores developed within two to three weeks. Although complete darkness is required for successful mating tests of some zygomycetes (e. g. Mucor spp.) [29], we determined that R. oryzae group isolates formed zygospores in partial light or in complete darkness. During mating, most zygomycetes form a straight line of zygospores at the interface between the two compatible strains, which has been reported previously for R. oryzae [30] (C. Skory, personal communication). However, we did not observe the typical straight line of zygospores and instead found them diffusely distributed across the plate within ca 4 to 5 cm of the inoculation points. The majority of the zygospores were observed on the side of the Petri plate where the (+) strain was growing.

Under the light microscope, initiation of individual zygospores between two conjugating hyphae could be observed. Conjugating hyphae were consistently shorter and thicker than vegetative hyphae, and were separated from vegetative hyphae by visible

<table>
<thead>
<tr>
<th>Collection #</th>
<th>Zygospores</th>
<th>Mating type</th>
<th>Origin and notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>R. oryzae s. s.</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CBS112.07</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>CBS127.08</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>CBS148.22</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>as R. tonkinensis, lactic acid</td>
</tr>
<tr>
<td>CBS257.28</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>as R. fumigatus, fermented food</td>
</tr>
<tr>
<td>CBS266.30</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>CBS346.36</td>
<td>yes</td>
<td>plus 1, 2</td>
<td>non pathogenic</td>
</tr>
<tr>
<td>CBS382.52</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>produces steroids</td>
</tr>
<tr>
<td>Duke66.02</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>Duke99-133</td>
<td>no</td>
<td>n. d.</td>
<td>human pathogen</td>
</tr>
<tr>
<td>Duke99-892</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL395</td>
<td>no</td>
<td>plus 2</td>
<td>Hildebrandt</td>
</tr>
<tr>
<td>NRRRL1897</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>grain</td>
</tr>
<tr>
<td>NRRRL2908</td>
<td>yes</td>
<td>plus 1, 2</td>
<td>Chinese yeasts</td>
</tr>
<tr>
<td>NRRRL142</td>
<td>no</td>
<td>minus 2</td>
<td>Chinese yeasts</td>
</tr>
<tr>
<td>NRRRL833</td>
<td>no</td>
<td>plus 2</td>
<td>produces 6-azaureidine</td>
</tr>
<tr>
<td>NRRRL834</td>
<td>no</td>
<td>minus 2</td>
<td>barley</td>
</tr>
<tr>
<td>NRRRL614</td>
<td>no</td>
<td>minus 2</td>
<td>parsnip, produces steroids</td>
</tr>
<tr>
<td>NRRRL10206</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL2125</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL21789</td>
<td>yes</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL28631</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL336</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>wine cake</td>
</tr>
<tr>
<td>NRRRL10884</td>
<td>no</td>
<td>minus 1</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL13142</td>
<td>no</td>
<td>minus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRL13440</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>R. delemar</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CBS329.47</td>
<td>yes</td>
<td>minus 1, 2</td>
<td>tempeh, produces pectinase</td>
</tr>
<tr>
<td>NRRRL1528</td>
<td>no</td>
<td>minus 2</td>
<td></td>
</tr>
<tr>
<td>NRRRL1546</td>
<td>yes</td>
<td>minus 1, 2</td>
<td></td>
</tr>
<tr>
<td>NRRRL1549</td>
<td>no</td>
<td>plus 2</td>
<td></td>
</tr>
<tr>
<td>NRRRL1551</td>
<td>no</td>
<td>minus 2</td>
<td></td>
</tr>
<tr>
<td>NRRRL1552</td>
<td>no</td>
<td>minus 2</td>
<td></td>
</tr>
<tr>
<td>NRRRL2005</td>
<td>no</td>
<td>plus 2</td>
<td>produces fumaric acid</td>
</tr>
<tr>
<td>NRRRL3562</td>
<td>no</td>
<td>plus 2</td>
<td>tempeh</td>
</tr>
<tr>
<td>NRRRL3563</td>
<td>no</td>
<td>plus 2</td>
<td>tempeh</td>
</tr>
<tr>
<td>NRRRL3098</td>
<td>no</td>
<td>minus 2</td>
<td>tapioca (tapai-ubi)</td>
</tr>
<tr>
<td>NRRRL21447</td>
<td>no</td>
<td>minus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>NRRRLA-16456</td>
<td>no</td>
<td>minus 2</td>
<td>human pathogen</td>
</tr>
<tr>
<td>RA99-880</td>
<td>no</td>
<td>plus 2</td>
<td>human pathogen</td>
</tr>
</tbody>
</table>

1determined in mating assays.
2determined by sequencing the sex locus.
3strains for which the sex locus and flanking genes have been sequenced.

type culture.
4tester strains used in mating assays.
n. d. – not determined.
doi:10.1371/journal.pone.0015273.t001
After zygosporangia formation, conjugating hyphae developed into asymmetric suspensor cells (Fig. 1) [3]. Zygospores were morphologically variable and ranged from 60–140 μm in diameter. Zygospore shape ranged from round to flat with stellate conical projections (Fig. S2A) and color ranged from reddish brown to dark brown. As in previous mating studies of Rhizopus oryzae s. l., we also observed large, central vacuoles inside many zygospores [21].

Contrary to published results for R. stolonifer [25,26], we were unable to stimulate germination of zygospores into germosporangia under any of the experimental conditions tested. Gentle crushing of exospor (outer zygospore wall) [31] resulted in the release of protoplast material, which developed into a vegetative mycelium (Fig. S2C). Fourteen single spore isolates were obtained using the described primers, we obtained DNA from these cultures as previously described for R. oryzae s. l. DNA was extracted from these cultures as previously described for standard isolates. Using the described primers, we obtained DNA sequences for the sex locus of these isolates and determined that all 14 single spore isolates had only the (−) mating type.

Structure of the mating locus in R. oryzae

Recent studies demonstrated that P. blakesleeanus and M. circinelloides have similar sex genes [16,27]. The sex locus and the flanking genes in both species consists of a 7–10 kb region that includes a gene encoding a high mobility group (HMG) that is flanked by a gene encoding a TP transporter (TPT) and a second gene coding for an RNA helicase. DNA sequences of P. blakesleeanus and M. circinelloides were used to screen the publicly available genome of R. oryzae and identify the sex locus in R. oryzae [16]. The sex locus of R. oryzae is located in a 13 kb region in Supercontig 1 (R. oryzae database, http://www.broadinstitute.org/annotation/genome/rhizopus_oryzae/MultiHome.html). It encodes proteins with 48% and 38% total amino acid identity with the (+) sex loci of P. blakesleeanus and M. circinelloides, respectively. Unlike P. blakesleeanus and M. circinelloides, which have sex and TPT genes in an inverted orientation [16,27], the sex and flanking genes of both (+) and (−) strains of R. oryzae are oriented in the same direction (Fig. 2).

We designed specific PCR primers to amplify the sex loci from five representative strains of R. oryzae; one of which contained the putative (+) mating type and four strains with the putative (−) mating type. DNA sequencing confirmed that the mating locus structure of R. oryzae is similar to those of P. blakesleeanus and M. circinelloides and consists of an HMG gene (SexP/SexM) flanked by TP transporter and RNA helicase genes (Table 2, Fig. 2). The mating genes (HMG, TPT, and RNA helicase) of the (+) strains of these species share 24% amino acid identity (49% similarity at the DNA level). The (−) alleles of these three species share 30% amino acid identity (92% similarity at the DNA level).

The RNA helicase gene is 4161–4191 bp long and includes nine introns of 40–150 bp each. This gene was annotated in the R. oryzae database as transcript RO3G_01291 in Supercontig 1. The TP transporter gene, which is not yet annotated in the R. oryzae database, is 1336 bp long and belongs to the EamA-like transporter family. We also identified the putative protein-coding gene [16] with an ORF that was located between the TPT gene and the HMG gene in both the (+) genome strain RA99-880 and the (−) tester strain CBS346.36. Parts of this gene are annotated in the R. oryzae database as transcripts RO3G_01290 (1482 bp), RO3G_01289 (1831 bp), and RO3G_01288 (625 bp). These transcripts were identified as an Ankyrin repeat region, a BTB/POZ region, and a third protein of unknown function, respectively. This three-part region BTB/Ankyrin/RCC1 (Fig. 2) [16] is absent in all of the (−) strains that we tested as well as the publicly available genomes of Phycomyces blakesleeanus and Mucor circinelloides [16,27]. The presence of BTB/Ankyrin/RCC1 accounts for the fact that the sex locus and flanking gene complexes from (−) strains are smaller (7 kb) than in the (+) strains (13.3 kb). Aside from BTB/Ankyrin/RCC1, the structure and direction of the flanking genes is similar in (−) and (+) strains.

Strain RA99-880, used to obtain the genome sequence of R. oryzae [32], did not mate with any of the isolates that we tested. This observation led us to question whether this isolate represents R. oryzae or a different Rhizopus species (Table 3). We compared the (+) mating alleles of RA99-880 to CBS346.36. We previously determined that CBS346.36 produced zygosporangia when mated with the holotype culture of R. oryzae CBS112.07, suggesting that they are likely conspecific. We determined that the HMG box region in the (+) strain CBS346.36 was 942 bp long, contained no introns, and shared only 93% DNA sequence identity with genome strain RA99-880.

In addition, we compared the structures of the HMG box region between (−) and (+) alleles. The (−) allele encodes a protein that is 180 amino acids long whereas the (+) allele encodes a protein that is 253 amino acids in length. Both (+) and (−) alleles share 23% nucleotide identity but both contain an identical amino acid motif at the 5′-end of their sequences: RPT[NAD][LY]. This motif is nearly identical to the motifs found in both the (+) and (−) strains of Phycomyces blakesleeanus and Mucor circinelloides.
of the TPT and RNA helicase genes are highly conserved. The orientations of these genes are the same in \( M. \text{circinelloides} \) and \( R. \text{oryzae} \). The orientations of sexP and sexM genes are the same in \( R. \text{oryzae} \). Relative to \( R. \text{oryzae} \), the TPT gene in \( M. \text{circinelloides} \) and both the TPT gene and sexP in \( P. \text{blakesleeanus} \) have an inverted orientation. The (+) allele of \( R. \text{oryzae} \) contains an additional gene between the TPT gene and sexP that is not found in the other two species or in the (−) allele of \( R. \text{oryzae} \). Black boxes show repetitive sequence tracts, arrows show gene orientation, and lines represent intergenic sequence.

doi:10.1371/journal.pone.0015273.g002

**Multilocus phylogenetic studies**

Comparative analysis of mating loci between two (+) and four (−) strains of \( R. \text{oryzae} \) revealed that two (+) strains, CBS346.36 and RA99-880, shared 93.3% amino acid identity. Although the DNA sequences from (−) strains were more conserved, we still detected significant differences between isolates: these four strains shared 98.4–100% amino acid identity. This genetic divergence between isolates of the same mating type prompted analysis of genetic diversity among multiple isolates of \( R. \text{oryzae} \).

To determine genetic relationships among isolates, we obtained partial sequences of the HMG gene from 57 strains using PCR primers specific for (+) and (−) mating alleles. Based on their DNA sequences, 26 strains belonged to the (+) type and 29 strains belonged to the (−) type (Table S3). These partial sequences were used for determining phylogenetic relationships among the strains.

Eleven haplotypes of the (+) HMG locus were identified, which were separated into two distinct groups on the ML tree (Fig. 3A). Fifteen different haplotypes of the (−) HMG locus were identified, which also were separated into two well-supported phylogenetic groups (Fig. 3B). We also obtained partial sequences from the two flanking genes, TPT and RNA helicase. Eighteen TPT and 16 RNA helicase haplotypes were detected among the 41 strains.

Gene trees for each locus strongly support two clades (Figs. S1C, D). Four additional gene regions commonly used in fungal systematics (mitochondrial small-subunit RNA (mtSSU), nuclear-encoded large subunit RNA (nLSU 28S), ITS1-5.8S-ITS2 region and \( RPB2 \)) were also sequenced from the complete set of 57 clinical and environmental strains (Table S3). MtSSU and rDNA 28S genes were too highly conserved to provide phylogenetic resolution within the \( R. \text{oryzae} \) complex (data not shown). In contrast, phylogenetic analysis of the more variable rDNA ITS1-5.8S-ITS2 and \( RPB2 \) regions revealed the same two well-supported clades detected by the analysis of the mating loci. Phylogenetic analysis of the concatenated data set containing all four loci (rDNA ITS1-5.8S-ITS2, \( RPB2 \), TPT and RNA helicase) also strongly supports two major clades (Fig. 3C, also Fig. S1A). These two clades correspond to two cryptic species, \( R. \text{oryzae} \) sensu stricto and \( R. \text{delemar} \), that had been previously recognized based on \( LDH \) genes and ITS sequencing [9].

Although separate and combined gene trees all support evidence for two cryptic species, phylogenetic relationships among strains within each species varied depending on which gene was used. This is also illustrated by the failure of consensus trees to resolve phylogenetic relationships within either cryptic species (Fig. 3D). A phylogenetic congruence test [33] revealed significant discordance among gene phylogenies attributable to within-species recombination among all 4 loci (partition homogeneity test, \( p = 0.001 \)). This scenario, whereby multiple gene trees consistently resolve the phylogeny between but not within species, is the basis for the Phylogenetic Concordance Species Concept advocated by Taylor et al. [34].

When we mapped results of the mating experiments to the ITS+\( RPB2+TPT+RNA \) helicase phylogeny (Fig. 3C) we determined that many isolates of \( R. \text{oryzae} \) s. s. were capable of producing zygospores when paired with isolates of the opposite mating type. Specifically, three (+) and eight (−) strains out of 33 \( R. \text{oryzae} \) s. s. isolates (11/33 or 33%) were capable of a successful mating reaction as defined by the production of zygospores. In contrast, most \( R. \text{delemar} \) strains were sterile. Only three of 21 \( R. \text{delemar} \) strains were capable of producing zygospores when paired...
with the (+) tester strain CBS346.36 (3/21 or 14%). However, none of the strains produced zygospores when paired with the (+) genome strain RA99-880 from the same clade.

Distribution of mating types within each cryptic species

To test if the distribution of the two mating types was the same in populations of *R. oryzae* and *R. delemar*, we used mating gene-specific PCR primers to assign mating types to 57 different strains. All strains possessed either a (+) or a (−) HMG mating allele (mating allele type could not be determined for two strains). As expected, (+) strains were only capable of mating with (−) strains and vice versa. In each species, the ratio of plus/minus mating types is close to 1:1, evidence that is consistent with a randomly mating population. Within *R. oryzae*, the ratio of +/− isolates was observed to be 14:18 (chi-square with one degree freedom = 0.5, two tailed p = 0.4795). For *R. delemar* this ratio of plus/minus was 12:11 (chi-square = 0.043, p = 0.8348). For all isolates combined (regardless of species) this ratio (26:29) is also not significantly different from 50:50 (chi-square = 0.164, p = 0.6568).

Asexual sporangiospores of *R. oryzae* and *R. delemar* are morphologically indistinguishable

When grown on agar media, all *R. oryzae* complex strains have similar growth patterns and growth rates. Actively growing colonies fill a 90 mm Petri dish in 2 to 3 days at room temperature (23°C). After 3 to 4 days, most isolates begin to produce sporangiospores (Fig. S2B), although some strains sporulate more slowly and may require 10 to 14 days.

In an effort to delimit *R. oryzae* from *R. delemar* isolates based on morphology, we measured the length and width of sporangiospores from 20 randomly chosen strains. The *R. oryzae* and *R. delemar* isolates we studied showed no observable differences in spore size (Fig. 4). Size differs considerably even for the spores produced by the same sporangium (Fig. 5), which reflects the different number of nuclei per spore. Binucleate sporangiospores are considerably larger than uninucleate ones (Fig. S2D). Our findings agree with previous studies [8] that both *R. oryzae* and *R. delemar* can have a wide and overlapping range of spore sizes. The length to width ratio remains approximately constant at 1:3.

Sporangial surface area was strain-specific for each of 12 randomly selected *R. oryzae* complex isolates (Fig. S3). However, we were unable to determine a specific pattern distinguishing sporangiospores of *R. oryzae* from *R. delemar*.

*Rhizopus oryzae* s. s. and *R. delemar* strains can be differentiated by PCR with primers specific to lactate dehydrogenase (*LDH*) genes

Abe et al. [10] demonstrated that *R. oryzae* contains two copies of *LDH*, *LDHA* and *LDHB*, whereas *R. delemar* has only a single

---

**Table 2.** Comparison of the number of genes, the length of individual genes, and the length of the entire gene complex in the sex loci of *P. blakesleeanus, M. circinelloides* and *R. oryzae* complex.

<table>
<thead>
<tr>
<th>Parts of mating type locus</th>
<th><em>P. blakesleeanus</em></th>
<th><em>M. circinelloides</em></th>
<th><em>R. oryzae</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>GenBank accession #</td>
<td>EU009462</td>
<td>EU009461</td>
<td>FJ009107</td>
</tr>
<tr>
<td>Total fragment length</td>
<td>9.97</td>
<td>7.63</td>
<td>6.88</td>
</tr>
<tr>
<td>Intergenic regions, total length</td>
<td>5.33</td>
<td>3.53</td>
<td>0.57</td>
</tr>
<tr>
<td>TPT (EamA-like transporter family)</td>
<td>289</td>
<td>-</td>
<td>276</td>
</tr>
<tr>
<td>BTB domain &amp; ankyrin repeat (RO3G_01289)</td>
<td>-</td>
<td>-</td>
<td>395</td>
</tr>
<tr>
<td>BTB/POZ domain (RO3G_01289)</td>
<td>-</td>
<td>-</td>
<td>649</td>
</tr>
<tr>
<td>Predicted protein, Rcc1 (RO3G_01290)</td>
<td>-</td>
<td>-</td>
<td>258</td>
</tr>
<tr>
<td>HMG (MATA box)</td>
<td>-</td>
<td>-</td>
<td>632</td>
</tr>
<tr>
<td>RNA helicase (RO3G_01291)</td>
<td>-</td>
<td>-</td>
<td>526</td>
</tr>
<tr>
<td>Repetitive elements</td>
<td>1</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Inverted repeats</td>
<td>-</td>
<td>-</td>
<td>1</td>
</tr>
<tr>
<td>Palindromes</td>
<td>-</td>
<td>-</td>
<td>3</td>
</tr>
</tbody>
</table>

1Inverted genes.
2Single ORF.
3Length of DNA shown in kilobases.
4Length of protein shown in amino acids.

**Table 3.** Comparison of the phylogenetic signal obtained from different genes using different methods for delimitation of the two cryptic species, *Rhizopus oryzae* s. s. and *Rhizopus delemar*; percent difference is depicted for both DNA (bp) and amino acids (aa).

<table>
<thead>
<tr>
<th>Locus</th>
<th>Length (bp)</th>
<th>% Difference (bp)</th>
<th>% Difference (aa)</th>
<th>ML Bootstrap Support</th>
</tr>
</thead>
<tbody>
<tr>
<td>mtSSU</td>
<td>370</td>
<td>-</td>
<td>n. a.</td>
<td>-</td>
</tr>
<tr>
<td>rDNA 28S</td>
<td>950</td>
<td>0.3</td>
<td>n. a.</td>
<td>-</td>
</tr>
<tr>
<td>rDNA ITS1-5.8S-ITS2</td>
<td>526</td>
<td>1.7</td>
<td>n. a.</td>
<td>83</td>
</tr>
<tr>
<td>RPB2</td>
<td>759</td>
<td>3</td>
<td>4.7</td>
<td>99</td>
</tr>
<tr>
<td>HMG (+)</td>
<td>466</td>
<td>3.7</td>
<td>4.5</td>
<td>100</td>
</tr>
<tr>
<td>HMG (−)</td>
<td>632</td>
<td>3.8</td>
<td>6.2</td>
<td>99</td>
</tr>
<tr>
<td>TPT</td>
<td>978</td>
<td>4.1</td>
<td>5.5</td>
<td>100</td>
</tr>
<tr>
<td>RNA helicase</td>
<td>764</td>
<td>5.1</td>
<td>7.2</td>
<td>100</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0015273.t002
doi:10.1371/journal.pone.0015273.t003
copy of this gene. We developed PCR primers that amplify portions of the LDHA and LDHB genes and tested them on 12 isolates from the R. oryzae complex, including six randomly selected isolates each of R. oryzae s. s. and R. delemar (Fig. 6A). All of the R. oryzae s. s. isolates produced an amplification product for both LDHA (ca 500 bp) and LDHB (ca 230 bp). In some cases, a nonspecific, higher molecular weight PCR product was also apparent. Sequencing of PCR products of R. delemar isolates confirmed it as the expected portion of LDHB. For R. delemar only LDHB products were amplified (Fig. 6B).

Figure 3. Maximum Likelihood phylogenies of the Rhizopus oryzae complex clearly delimit the two cryptic species, Rhizopus oryzae s. s. and Rhizopus delemar. (A) sexP alleles, (B) sexM alleles, (C) concatenated phylogram of rDNA ITS1-5.8S-ITS2, RPB2, TPT gene, and RNA helicase gene, and (D) four-gene strict consensus tree. Analysis included a total of 458 (+), 635 (−) and 3064 (MLS) nucleotide characters. ML bootstrap proportions higher than 70 are shown above the nodes. Asterisks (*) indicate strains that produced zygospores. Filled circles indicate a single R. delemar strain, CBS329.47, closest to the ancestral state.

doi:10.1371/journal.pone.0015273.g003
Discussion

Structure of the sex type locus in the *R. oryzae* complex

We examined the structure of the putative sex type locus in six representative strains of *R. oryzae* and demonstrate that *R. oryzae* possesses a sex type locus that is homologous to those of other zygomycetes and also of microsporidians. This gene cluster consists of an HMG domain gene flanked by RNA helicase and TP transporter genes [16]. The characterization of *R. oryzae* sex genes constitutes the third representative of the Mucoromycotina (Zygomycota) for which the mating gene structure is known. When compared with the recently described sex loci of *P. blakesleeanus* and *M. circinelloides*, the (+) allele of the *R. oryzae* sex locus was significantly larger due to the presence of an additional putative ORF. However, this inserted gene is absent in the (−) allele of the locus. Unlike the sex locus of *P. blakesleeanus* and *M. circinelloides*, which contain TPT and HMG genes in inverted orientations, all genes in the *R. oryzae* sex type locus were positioned in the same orientation (Fig. 2). The overall similarity between the sex type locus of *R. oryzae* and other zygomycetes was significant, suggesting that this locus is involved in regulation of the mating process [28]. Further functional genetic analysis is necessary to confirm this hypothesis.

Cryptic speciation in the *Rhizopus oryzae* complex

Our phylogenetic analyses confirm the previous observations by Abe et al. [10,35] that *R. oryzae* s. l. can be subdivided into two cryptic species groups, designated *R. oryzae* s. s. and *R. delemar*. We
confirmed that the genome strain RA99-880, once thought to belong to *R. oryzae* s. s., is actually a member of *R. delemar* [32]. Genetic isolation between these two cryptic species was evident in the strongly supported four-gene phylogeny and the genealogies of both the (+) and (−) alleles of the HMG genes (Fig. 3). The higher number of amino acid and nucleotide changes in the sex genes relative to the “standard” genes (Table 3) suggests that the sex genes may prove useful in resolving phylogenetic questions in other zygomycetes and perhaps in other fungi.

Saito et al. [14] were also able to distinguish *R. oryzae* s. s. and *R. delemar* based on rDNA phylogeny and physiological differences in acid production. They confirmed the presence of two distinctive physiological groups: isolates that produce lactic acid and have two copies of the *LDH* gene (*R. oryzae* s. s.) and isolates that produce both fumaric and malic acids but have only one *LDHB* (*R. delemar*). We designed primers for the *LDH* genes and confirmed a similar pattern in the 57 isolates we studied. It has been suggested that *R. oryzae*’s ability to ferment various organic substrates may explain why this species was domesticated and used to process various types of Asian fermented foods [4,11]. Lactate dehydrogenases (LDH) are important for these enzymatic processes [2,4,11,36,37].

Despite the fact that *R. oryzae* s. s. and *R. delemar* can be resolved by multi-locus phylogenetic analyses and physiological differences in acid production, morphological examinations and molecular analyses of more conservative loci indicate that these two cryptic species are very close relatives. For example, sequences of the mtSSU and 28S rDNA genes were too highly conserved to provide species are very close relatives. For example, sequences of the mtSSU and 28S rDNA genes were too highly conserved to provide any phylogenetic resolution within the *R. oryzae* complex. Similarly, analyses of secondary structure in the ITS2 region indicates that there are no complementary base pair changes (CBC) between *R. oryzae* s. s. and *R. delemar*, whereas other species in the genus *Rhizopus* are quite different (Table 4). Furthermore, isolates of *R. oryzae* s. s. and *R. delemar* exhibited similar colony morphology, growth characteristics, and spore morphology. Although sporangiospore size is of foremost importance in the classification of the genus *Rhizopus* [38,39], spore size measurements were insufficient to distinguish *R. oryzae* from *R. delemar*.

### Sexual reproduction of *R. oryzae* and other zygomycetes in the laboratory and in nature

Fungi employ a variety of reproductive modes, in the lab as well as in nature. Whereas some fungal species appear to be obligate sexual organisms (e.g. [40,41]) and others appear to be strictly asexual [42,43,44], the majority of fungi apparently have a mixed mating system that includes both sexual and asexual reproduction (e.g. [45,46,47]). Although earlier studies suggested a predominantly asexual mode of reproduction in *R. oryzae* [3,4], analysis of sequence variation in multiple loci suggests that this species complex comprises two cryptic species and both exhibit a sexual mode of reproduction.

Zygospore formation is considered a hallmark of sexual reproduction. However, only 10% of described Mucoromycotina species are known to produce zygospores, either under natural conditions or in pure culture [48,49]. Despite relatively frequent formation of zygospores in closely related heterothallic (*R. stolonifer, R. microsporus*) and homothallic (*R. sexualis, R. homothallicus*) species, only a few reports have described zygospore production between isolates of the *R. oryzae* complex [3,4,21,25,26]. In our study, 40% of the isolates of *R. oryzae* s. s. were capable of producing zygospores in the laboratory when paired with isolates of the opposite mating type. The ability to produce zygospores with a suitable mating partner seems to be characteristic of almost all subgroups of the *R. oryzae* s. s. clade (Fig. 3).

Most strains producing zygospores in our study belong to the *R. oryzae* s. s. clade (Table S3). Although most isolates of *R. oryzae* s. s. were capable of producing zygospores, we observed that their morphology was more similar to the azygospores observed in other zygomycetes. Zygospores of *R. oryzae* possess asymmetric suspensors of unequal size, a feature associated with azygospore production in *R. stolonifer* and *R. microsporus* [3,6,18,19]. Single asymmetric suspensors have been reported in different varieties of *R. microsporus* [4], and in distantly related species from the genera *Absidia* and *Zygorhynchus* [50]. In the genus *Rhizopus*, zygospore formation may not be a useful character for delimiting species boundaries because many *Rhizopus* isolates form zygospores even when their mating partner belongs to a different species [51].

In our study, *R. oryzae* zygospores did not germinate into germosporangia [52] under any of the tested experimental conditions. When zygospores were mechanically disrupted, the multinuclear proplastid developed into a vegetative mycelium that contained nuclei of only the (−) mating type, indicating that meiosis did not occur. If meiosis had occurred, we would instead expect equal proportions of (+) and (−) mating types. This result suggests that mating and recombination might not occur under laboratory conditions.

In contrast, zygospores of the closely related species *R. stolonifer* can germinate in two different ways. When zygospores are germinated prior to maturation, they form a vegetative mycelium that is either (+) or (−). When mature zygospores are germinated, they form a germosporangium that produces spores with equal proportions of (+) and (−) mating types [25,26,49]. Germination of zygospores into vegetative mycelia is also known in distantly related species in the genus *Zygorhynchus* [21].

Although we did not directly detect successful mating and progeny development in the laboratory, we did observe nearly equal frequency of (+) and (−) mating types across both cryptic species within the *R. oryzae* species complex (Table S3). This provides strong indirect evidence for sexual recombination in *R. oryzae*. Analysis of multiple gene phylogenies provides further evidence for outcrossing. Phylogenetic analyses of individual and combined loci consistently support two well-defined sibling species, *R. oryzae* s. s. and *R. delemar*. Within each species, however, discordant gene trees suggest that both species are inherently sexual, with an outcrossing mode of reproduction (Fig. S1, supplementary materials). If populations of each species were clonal, we would expect independent gene trees to be concordant [33]. In this study, we consistently observed conflict between gene partitions within both species (ph test, p<.001), suggesting

### Table 4. Analysis of similarity between *Rhizopus oryzae* s. s. and other *Rhizopus* species in ribosomal DNA sequences (ITS1-5.85-ITS2, 28S) and predicted compensatory base changes (CBC) for the ITS2 region.

<table>
<thead>
<tr>
<th>Species</th>
<th>rDNA ITS1-5.85-ITS2</th>
<th>rDNA 28S</th>
<th>CBC</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>R. delemar</em></td>
<td>98.3% (HQ435103)</td>
<td>99.7% (HQ435039)</td>
<td>0</td>
</tr>
<tr>
<td><em>R. microsporus</em></td>
<td>65% (HQ435072)</td>
<td>95% (HQ435046)</td>
<td>2</td>
</tr>
<tr>
<td><em>R. homothallicus</em></td>
<td>68% (EU991016)</td>
<td>94% (DQ641324)</td>
<td>2</td>
</tr>
<tr>
<td><em>R. zygosporus</em></td>
<td>43% (DQ641314)</td>
<td>93% (DQ646599)</td>
<td>2</td>
</tr>
<tr>
<td><em>R. caespitosus</em></td>
<td>43% (DQ641325)</td>
<td>93% (DQ646604)</td>
<td>2</td>
</tr>
<tr>
<td><em>R. schipperae</em></td>
<td>43% (DQ641323)</td>
<td>92% (DQ646606)</td>
<td>2</td>
</tr>
<tr>
<td><em>R. sexualis</em></td>
<td>55% (DQ641322)</td>
<td>89% (DQ646592)</td>
<td>8</td>
</tr>
<tr>
<td><em>R. stolonifer</em></td>
<td>39% (FN401529)</td>
<td>89% (DQ646595)</td>
<td>6</td>
</tr>
</tbody>
</table>

doi:10.1371/journal.pone.0015273.t004
recombination between loci. Together with equal distribution of sexes, our results strongly support the hypothesis that genetic recombination is occurring within both *R. oryzae* s. s. and *R. delemar*. Further studies employing population-level sampling, along with genome sequencing of *R. oryzae* s. s. will be necessary to determine the extent and nature of recombination in both species.

Mucormycosis is an emerging infectious disease [33] and is increasingly reported as a cause of fungal infection in patients with impaired immunity [17]. Isolates from the *Cryptococcus neoformans* complex are the most common zygomycetes infecting humans [54,55]. Our phylogenetic studies also suggest that the two cryptic species, *R. delemar* and *R. oryzae*, may differ in relative virulence. It is also possible that *R. delemar* and *R. oryzae* may differ in susceptibility to antifungal drugs [55,56]. In the pathogenic basidiomycete, *Cryptococcus neoformans*, the MAT locus is linked to virulence [57,58]. It is possible that the sex locus might be similarly involved in the pathogenesis of *Rhizopus* species.

**Conclusion**

Our data support the hypothesis that the genetic machinery for sexual reproduction is conserved among multiple genera of Mucoromycotina. *Rhizopus oryzae* is one of the most economically important members of this group. Our multi-locus phylogenetic analyses support the existence of two cryptic species: *R. delemar* and *R. oryzae*. Notably, the recently sequenced genome isolate is a member of *R. delemar*, so it would be ideal to also sequence a representative genome of *R. oryzae* s. s. We demonstrate that both species have the potential for sexual reproduction, and although our mating studies suggest that sexual reproduction is infrequent, strains of both mating types are equally abundant in both species.

**Materials and Methods**

**Cultivation of fungal strains**

Strains of *Rhizopus oryzae* s. l. were obtained from different culture collections: American Type Culture Collection (ATCC), Duke University, Fungal Genetics Stock Center (FGSC), Agricultural Research Service Culture Collection (NRRL) in USA, Centraalbureau voor Schimmelcultures (CBS) in the Netherlands and Friedrich-Schiller University Fungal Reference Center in Germany (Table S3). Replicate isolates of several mating type tester strains first described for *R. oryzae* by Schipper [3] were included. Strains were grown on 10% Potato-Dextrose Agar (PDA) (NEOGEN, Lansing, MI, USA), in tubes and 90 mm Petri plates at room temperature.

**Light microscopy and fungal morphology**

Wet mounts of asexually sporulating isolates were examined at 100–400× under an Olympus CX31 light microscope equipped with an ocular micrometer (Olympus America Inc., Center Valley, PA, USA). To test whether morphology could be used to differentiate genetically related strains within the *R. oryzae* complex we made 100 measurements of sporangiospore width and length from 20 randomly chosen strains (Fig. 4). We then analyzed the morphology and size of *R. oryzae* s. l. spores using multivariate discriminate analysis and cluster analysis [59,60,61]. The following statistical parameters were examined for asexual sporangiospore dimensions: average (*M*), variance (*D*), standard deviation (*σ*), and standard error (±*m*). All statistical analyses were conducted in the STATISTICA 6.0 software package (StatSoft Inc., Tulsa, OK, USA).

**Mating assays**

Mating tests were conducted in 4.5 or 9 cm diameter Petri plates with several types of standard agar growth media: PDA, Malt Agar (MA) (VWR International, Bristol, CT, USA), Glucose Peptone Yeast Extract Agar (GPYA) (HiMedia Laboratories Pvt. Ltd., Mumbai, India), Sabouraud agar [62], and water agar [52]. Mating tests were conducted at three different nutrient levels; 10%, 50%, or 100% of the level recommended by the manufacturer to test the affect of nutrients on zygospore germination. Petri plates were either inoculated with 1×1 cm agar pieces taken from the growing edge of the colony, or with a 20 μl suspension of lyophilized tissue. In each mating test inocula from two isolates were placed 1 cm from each other on a fresh Petri plate. Cultures were then incubated in either complete darkness or exposed to light and they were either sealed or not sealed with Parafilm (Pechiney Plastic Packaging, Menasha, WI, USA) at room temperature according to Heselton and Rogers [29]. Development of conjugating hyphae and zygospores was observed at 100–400× under an Olympus CX31 light microscope (Olympus America Inc., Center Valley PA, USA) or at 200–3500× with a JSM-5900LV scanning electron microscope (SEM) (JEOL U.S.A., Peabody MA, USA). Observations for mating were conducted several times each week for a period of 6–8 weeks. For mating crosses that produced zygospores, we attempted to stimulate germination following traditional protocols [29]. We also tested whether it was possible to grow ungerminated zygospores by crushing them with forceps and then incubating them on nutrient agar.

**DNA extraction, amplification and sequencing**

Prior to DNA isolation cultures were incubated for 8 to 20 hours at room temperature in 50 ml of Potato Dextrose broth (NEOGEN, Lansing, MI, USA) and were then filtered through a sterilized Mira cloth [36]. Mycelium was then lyophilized for 1 to 2 days and ground in liquid nitrogen with a mortar and pestle or glass beads.

DNA was extracted following the CTAB extraction technique [63]. PCR mixtures with a total volume of 25 μl per sample were prepared according to the protocol supplied by TaKaRa (TaKaRa Bio Inc., Otsu, Shiga, Japan). PCR was performed according to the basic protocols outlined by White et al. [64] for all genes except where specifically noted below. The internal transcribed spacer region of rDNA (ITS1-5.8S-ITS2) was amplified with primers ITS1 and ITS4 [64]. Partial large subunit rDNA 28S was amplified using the primers LROR [65] and LR5 [66]. The mitochondrial SSU was amplified with the primer set mtSSU1_f and mtSSU2_r [67]. The genome of *R. oryzae* has two non-identical copies of *RPB2* located on different supercontigs. Accordingly, it was not possible to use the coding domains 5–7, which are located in the central part of the gene and have traditionally been used in fungal phylogenetics. To overcome this problem, we obtained DNA sequences of coding domains 1–3, which are unique to the *RPB2* copy located on Supercontig 10. To amplify the 5′ end (domain 1, intron 1 and part of domain 2) of the RNA Polymerase II subunit 2 gene (*RPB2*), we designed three new primers (*RhsRPB2f*, *RhsRPB2r* and *RhsRPB2xr* – Table S1) using the *RPB2* sequence from the *R. oryzae* genome database. For *RPB2* a “touchdown” PCR protocol was used as described by Don et al. [68].

All PCR-amplified fragments were separated by electrophoresis on a 1.5% agarose gel stained with SYBR® Safe and visualized with a UV transilluminator. PCR products were purified with Qiagen Quick-Clean columns (Qiagen Inc., Valencia, CA, USA) or with ExoAP enzymes [69]. PCR products were then sequenced using amplification primers and Big Dye chemistry version 3.1 (Applied Biosystems Inc., Foster City, CA, USA), and the DNA sequences were run on an ABI3700 DNA sequencer (Applied Biosystems Inc., Foster City, CA, USA).
Biosystems Inc., Foster City, CA). DNA sequences generated for this study were edited in Sequencher 4.0 (Gene Codes, Ann Arbor, MI, USA), and are available in GenBank (Table S2).

**Phylogenetic analysis**

We characterized rDNA 28S and ITS1-5.8S-ITS2, mtSSU, HMG, RPB2, TPT, and RNA helicase gene for 41 isolates (Table 3). We also examined the phylogenetic pattern of the “plus” and “minus” mating alleles to see whether these regions could be used to distinguish between different lineages from the *R. oryzae* complex. First, we compared the phylogenetic signal of the “plus” and “minus” alleles to other genes that have traditionally been used for phylogenetic analyses of fungi (rDNA 28S and ITS1-5.8S-ITS2, RPB2). Next, we compared the phylogenetic signal of the “plus” allele with the “minus” allele to see if inferences from each of these alleles were similar and which region was more informative. Phylogenetic relationships were determined by Maximum Likelihood (ML) and phylogenetic support was assessed for each gene by bootstrap analyses (ML) using PAUP* 4.0a109 [70]. Using sequences of DNA ITS1-5.8S-ITS2, RPB2, TPT, and the RNA helicase genes, the maximum likelihood approach was applied to generate a four genes tree for all of the 41 studied isolates. The partition homogeneity test was run on PAUP* 4.0a109 using a parsimony criterion for four genes and 1000 replicates [70].

After characterizing the sex genes of several *R. oryzae* isolates, we compared them to the previously characterized sex alleles of *Phycomyces blakesleeanus* (EU009461, EU009462) and *Macrocordyloides* (FJ009106, FJ009107). Nucleotide and amino acid similarity of HMG, TPT and RNA helicase gene/protein alignments was examined in Multalin [71] and ClustalW2 [72].

To generate a dataset of *Rhizopus* reference taxa, additional rDNA 28S and ITS1-5.8S-ITS2 sequences from identified *Rhizopus* species with accurate nomenclature were either downloaded from GenBank or generated in this study (Table 4) [73]. Compensatory base changes in the ITS2 region for *Rhizopus* species were calculated using the ITS2 Database and the programs 4Sale and CBCAnalyzer [74,75,76,77]. We did not detect any compensatory base changes between *R. oryzae* and *R. delemar* in the ITS2 region (Table 4).

All sequences were initially aligned in ClustalX [78]. Alignments were manually adjusted and ambiguous regions were excluded from the alignments using Mesquite 2.5 [79]. sex loci and other genes of the strains of interest were aligned and compared using Sequencher 4.0 (Gene Codes, Ann Arbor, MI, USA), Multalin [71] and ClustalW2 [72]. Repetitive sequences in “plus” and “minus” alleles of the sex locus were determined manually and using the Tandem Repeats Finder [80]. Inverted and palindromic repeats were identified using tools available at the REPEATS, SECONDARY STRUCTURE & MELTING TEMPERATURE web site (http://emboss.bioinformatics.nl/cgi-bin/emboss/). Directionality of the genes in the sex loci was determined in ORF Finder (www.ncbi.nlm.nih.gov/orffinder/) and displayed with a dot plot.

**Analysis of mating types**

Primers for the TPT and RNA helicase flanking genes were designed using Primer3 [81] from the *R. oryzae* database sequences. PCR reactions using these primers were conducted using the TaKaRa protocol (TaKaRa Bio Inc., Otsu, Shiga, Japan). Products were isolated and purified using a gel extraction kit (Qiagen GmbH, Hilden, Germany), cloned with a TOPO TA kit (Invitrogen, Carlsbad, CA, USA), and sequenced as described above. New primers were subsequently designed at the end of the newly sequenced region and the process was repeated by “primer walking” to obtain DNA sequences between the flanking genes. A total of fifty-seven primers were employed to sequence the entire length of the *Rhizopus sex* locus, including the flanking genes (Table S1). The entire sex locus and the two flanking regions were sequenced for five strains (Table 2) and these data were used to design primer sets for direct amplification of the mating loci: SgcneCORE_for and SgcneCORE_rev for the “minus” allele and Plus1_f and Plus1_r for the “plus” allele (Table S1). PCR with the SgcneCORE primer set produced a fragment of 780–800 bp whereas PCR with the Plus1 primer set produced a fragment of 465–480 bp. To avoid formation of secondary structures during amplification of intergenic regions of the sex locus, 1% DMSO was added to both PCR and sequencing mixtures [82].

**Use of the Lactate Dehydrogenase (LDH) gene to differentiate *R. oryzae sensu stricto* from *R. delemar***

Lactate dehydrogenase genes encode hydrolytic enzymes that enable *R. oryzae* to grow in decaying organic matter rich complex carbohydrates [30]. Previous research has shown that differences in the LDH genes can differentiate *R. delemar* from *R. oryzae* s. s. *Rhizopus delemar* has only one copy of the gene (LDHB) whereas *R. oryzae* s. s. has two copies (LDHA and LDHB) [11,14] (*Rhizopus oryzae* database). The sequences of these two genes are almost identical except that LDHA has a longer 3' ORF. Although we did not include this locus in our phylogenetic analyses, we developed a PCR-based method to efficiently distinguish between *R. oryzae* s. s. and *R. delemar* using a combination of one forward and two reverse primers for the LDH gene. The forward primer JOHE22917 anneals to both genes. The reverse primer JOHE22918 anneals to both LDHA and LDHB whereas the other reverse primer JOHE22919 recognizes only LDHA (Fig. 6A). When PCR is performed with all three primers, *R. oryzae* s. s. isolates yield two fragments of different sizes: one PCR product for LDHA and another for LDHB. Isolates of *R. delemar* yield only one PCR product, however. We randomly selected six strains each of *R. oryzae* s. s. and *R. delemar* to test the LDH PCR assay. Results were compared with phylogenetic analyses used to distinguish these species.

**Supporting Information**

**Figure S1** Maximum Likelihood phylogeny for rDNA ITS1-5.8S-ITS2 (A), RPB2 (B), TPT (C) and RNA helicase (D) genes. Analysis included a total of 566 (rDNA ITS1-5.8S-ITS2), 757 (RPB2), 978 (TPT) and 764 (RNA helicase) nucleotide characters. ML bootstrap proportions higher than 70 are shown above the nodes. Group * includes ITS sequences AB097299, AB181316-AB181330 of *Rhizopus delemar*; group ** includes the ITS sequences AB181303-AB181309, AB181311-AB181315, AB097334 of *Rhizopus oryzae* [10]. T gene. T ‐ type culture of *R. oryzae* s. s., T' indicates a strain with an rDNA ITS1-5.8S-ITS2 sequence that is identical to the type culture of *R. delemar* (CBS120.12).

(TIF)

**Figure S2** Sexual (zygospores) and asexual (sporangiospores) spores of *Rhizopus oryzae*. (A) Electron micrograph of a cross between *R. oryzae* strains CBS346.36 × CBS110.17 showing zygospores [black arrow heads], Scale bar = 50 μm. (B) Asexual sporangium of *R. delemar* strain NRRL3563 without sporangium wall. Scale bar = 20 μm. (C) Germinating of zygospore’s protoplast (white arrow) into vegetative mycelium after crushing of lateral sporangium wall (black arrows). Scale bar = 50 μm. (D) Different size of uni- (black arrow) and binucleate (white arrow)
References


Acknowledgments

We thank Kerry O’Donnel, Wiley Shell, Ashraf Ibrahim and Andrej Gregory for R. oryzae isolates; Tom Mitchell, François Latzoon, John Shaw, William Colquhoun, Bernie Ball, Sandra Boles, Kerstin Hofmann, Tami McDonald, Kathleen Miglia, Katalin Molnar, Heath O’Brien, Hannah Reynolds, Bernadette O’Reilly, Marianela Rodrigues-Carrers, Peter Szovenyi, Kerstin Voigt, Niel Terenteyeva, Oleksandr Savytskiy, Niel Garrett and Grit Walter for advice on experimental work, figure design, statistical and data analyses; Iryna Duldka, Richard Humber and Christopher Spencer for experimental ideas and morphological information on R. oryzae; Mary Berbee, Tim James, Asia Buchalo, Reinhold Poder,Blanka Shaw, Jason Jackson, Sarah Jackson and Cathie Aime for essential discussions; Lisa Bukovnik for sequence analysis; Sheri Frank for reagent and culture supply; Jolanta Miadlikovska, Bernie Ball and Suzanne Joneson for help with primer design and sequence analysis; Valerie Knowlton for SEM assistance, and Scarlett Geunes-Boyer for considerable text improvement.

Author Contributions

Conceived and designed the experiments: AG SCL RV JH. Performed the experiments: AG SCL APL TMP GB. Analyzed the data: AG SCL APL MES GB TMP IMA JH RV. Contributed reagents/materials/analysis tools: JH RV. Wrote the paper: AP SCL JH RV.