Modulating Unimolecular Charge Transfer by Exciting Bridge Vibrations

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Nonadiabatic electron transfer (ET) measures the rate of electron tunneling, often facilitated by intervening covalent and nonbonded interactions.1,4 Bridge structure therefore influences the ET kinetics, and bridge thermal fluctuations are predicted to modulate the tunneling propensity.4,5 Structure defines the coupling pathways, and thermal fluctuations enable the system to find configurations that enhance the interaction strength.5 An open and crucial question is whether or not bridge motion can be manipulated (driven) by an external field to control pathway interactions and ET kinetics. Here, we show that mid-IR driving of bridge vibrations produces ET kinetic slowing of a photoinduced charge separation reaction.

Recent theoretical analysis indicates that ET kinetics can be changed by controlling the coherence of inelastic tunneling pathway interferences in a molecular analogue of the double-slit experiment.1,4,7 In systems with two interfering ET pathways, the excitation of a pathway-specific bridge vibration, which may induce electron-vibration energy exchange, labels the ET pathway and therefore modifies pathway interferences.1,4,7 Affecting ET rates using mid-IR radiation3 is generally attractive because ultrashort laser pulses offer subpicosecond perturbation, and radiation in the mid-IR is chemically innocent. In addition to inelastic tunneling, excitation of bridge vibrations can perturb elastic-tunneling kinetics. The donor-acceptor (DA) coupling may be modulated by exciting a bridge vibrational mode without electron-phonon energy exchange. Related ideas for the control of currents in molecular wires are being addressed in the context of molecular electronics and inelastic-tunneling spectroscopy.8

Here, we report the first real-time observation of ET rate modulation by mid-IR excitation in a donor-bridge-acceptor (DBA) ensemble.9 This ensemble consists of an anthracene-derived acceptor linked to a dimethylaniline-containing donor by guanosine-cytidine (GC) hydrogen bonding (Figure 1A). The ET is probed in a 3-pulse experiment, performed with a sequence of UV, mid-IR, and visible pulses (Figure 1B), each of ca. 100 fs duration. The first pulse at 400 nm creates the acceptor-localized electronic excited state (ES) that then “captures” an electron from the donor with an ET rate constant, $k_{CS}$, of ca. (30 ps)$^{-1}$.9 After a small time delay, $\tau$, the second pulse centered at 1670 cm$^{-1}$ and $\sim 120$ cm$^{-1}$ in width is applied, targeting vibrational modes in DBA labeled in Figure 1A. The third pulse in the visible spectral region probes the sample absorbance as a function of the probe’s delay time, $T$ (Figure 1B).

The absorbance changes in the 3-pulse measurements were calculated using the equation $\Delta \text{Abs} = D_R - D = \log(I/I_0)$, where $D_R$ and $D$ are the optical densities and $I_R$ and $I$ are the probe signals with the IR pump on and off, respectively. Since the mid-IR pulses were chopped at half of the laser repetition rate, and the two consecutively recorded spectra were processed to obtain $\Delta$abs, the 3-pulse measurements are sensitive exclusively to changes in the sample induced by vibrational excitation, rather than by the ET process itself.

The measurements were performed in an all-Teflon flow cell with an optical path length of 130 μm at 23.5 ± 0.3 °C in a nitrogen atmosphere. To minimize the amount of unbound acceptor in the sample ($K_{assoc} = 3.8 \times 10^4$ M$^{-1}$), all time-resolved measurements were performed in mixtures with a 5-fold molar excess of the donor (75 mM) over the acceptor (15 mM).

Figure 1C shows the UV–vis absorption spectrum of the donor, acceptor, and DBA samples recorded at the same molar concentration. At wavelengths above 280 nm, the spectrum of DBA is the sum of the spectra of the components, indicating weak interaction of the electronic states of the donor and acceptor units. In contrast, the IR spectrum of DBA is not the simple sum of contributions from the two component parts (Figure 1D), clearly demonstrating GC association.

UV-pump/vis-probe spectroscopy was used to measure the transient spectra of the excited and charge separated states of DBA and of the acceptor. The transient spectra of the excited state (ES) and of the charge-separated state (CS) for DBA, measured at a 3

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Figure 1. (A) Structure of the DBA complex where R = 2’,3’,5’-tri-O-(tert-butyldimethylsilyl)ribofuranosidyl. The groups participating most strongly in vibrational modes in the 1630–1700 cm$^{-1}$ range are highlighted. (B) Pulse sequence used in the 3-pulse experiments. Linear UV–vis (C) and mid-IR (D) absorption spectra of the donor and acceptor samples in dichloromethane, both at 20 mM, and their 1:1 (v:v) mixture (DBA) multiplied by a factor of 2. Four DFT-computed vibrational transitions of DBA are shown with green bars. Note that the DFT computed frequencies were scaled down with a factor of 0.948.

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and 77.4 ps probe delay, respectively (Figure 2A), were found to be in good agreement with the previous data.\textsuperscript{9}

In the 3-pulse measurements, the mid-IR pulses were targeting two strong absorptions of DBA at 1650 and 1689 cm\(^{-1}\) (Figure 1D). DFT calculations and normal-mode analysis\textsuperscript{10} lead us to assign the peak at 1689 cm\(^{-1}\) to an asymmetric stretch of the two carbonyl groups. Note that the DFT computed frequencies were normalized to the absorption peak at 1689 cm\(^{-1}\) by scaling down by a factor of 0.948. The peak at 1650 cm\(^{-1}\) has several contributions, including a relatively weak symmetric stretch of the carbonyl groups (computed at 1659 cm\(^{-1}\)) and in-plane bending modes of the two NH\(_2\) groups (computed at 1640 and 1623 cm\(^{-1}\), Figure 1C, green bars). The CO symmetric stretching mode is too weak to explain the absorbance at 1650 cm\(^{-1}\); in fact, the peak is dominated by NH\(_2\) in-plane bending modes. It is important to appreciate that all vibrational modes with frequencies between 1630 and 1700 cm\(^{-1}\) are assigned to groups in the G–C linkage.

Figure 2A shows the 3-pulse transient spectra for DBA measured at time delays \(T\). Interestingly, the largest transients are observed at small time delays, \(T\). To illustrate the main expectations in the 3-pulse experiments (see text), (D) 3-Pulse transient kinetics for DBA and for the acceptor (inset) measured at 578.8 and 563.6 nm, respectively. Exponential fits are shown in red. The CS population difference \(\Delta CS\) is related to \(\Delta \lambda\) by \(\Delta \lambda = \epsilon(\lambda) / \Delta CS\).

**Figure 2.** 3-Pulse transient spectra for DBA (A) and the acceptor-only sample (C), measured at \(t = 2.7\) ps, and different delays, \(T\), indicated. For the acceptor sample (C) the vibronic coupling is the only source of the signal. In contrast, DBA transients have another contribution that can be seen most clearly at later delays, where the transient spectrum matches that of the charge-separated state (CS). To emphasize the similarity, the excited (ES) and charge-separated (CS) state spectra were multiplied by \(-0.02\) (DBA) and 0.01 (acceptor). (B) Simulated kinetic traces illustrating the main expectations in the 3-pulse experiments (see text). (D) 3-Pulse transient kinetics for DBA and for the acceptor (inset) measured at 578.8 and 563.6 nm, respectively. Exponential fits are shown in red. The CS population difference \(\Delta CS\) is related to \(\Delta \lambda\) by \(\Delta \lambda = \epsilon(\lambda) / \Delta CS\).
subject to vibrational excitation. The 3-pulse transient spectrum (Δ(abs)) averaged over T of 75–80 ps has an amplitude that is 1.8% of that observed for the CS spectrum (2-pulse UV/vis) measured under the same conditions. The negative sign of the transient spectra indicates that less CS state was produced in the presence of the IR excitation. Note that the observed modulation of the CS yield is not associated with a trivial temperature increase caused by vibrational relaxation of the initially excited modes; preliminary data show that the rate of CS increases at higher temperatures. This temperature effect is obviously different from what is observed under the conditions of vibrational excitation.

A simple kinetic scheme was used to model the results (Figure 3A). E, CS, and G denote the excited, charge-separated, and ground electronic states, respectively; V indicates the presence of high-frequency vibrational excitation of the bridging species. While the decrease of the CS yield can be explained by either a reduction of the CS rate (k(cs)) and/or an increase in the rate of (mostly) nonradiative decay (k₀), we assume that k(cs) is affected by vibrational excitation. The modeling showed that the shape of the ΔCS transient kinetics, which is a difference in the amount of the CS state accumulated with and without IR excitation, is very similar to the quantum yield of charge separation (QYCS) for CS, giving a k(cs)(V) of (70–107 ps)^{-1} for a quantum yield ranging from 0.5 to 0.9. The cooling time for the modes affecting the CS reaction (k₀) was taken at 10.6 ps, which is the cooling time of high-frequency modes under the same conditions. The negative sign of the transient spectra of Δ(abs) averaged over 0.2. Interestingly, the value of k(cs)(V) obtained from the modeling is found to vary moderately as a function of QYCS, giving a k(cs)(V) of (70–107 ps)^{-1} for a quantum yield ranging from 0.5 to 0.9.

Several mechanisms can be proposed to account for the observed decrease of the CS yield upon IR excitation. One mechanism includes a dynamic loosening of the H-bonds by high-frequency vibrational excitation. For example, the H-atom displacement of the NH₂ of guanosine involved in the in-plane bending motion (1622 cm⁻¹) only elongates the H-bond to the carbonyl oxygen of cytidine; it never shortens it. The amplitude of the normal mode at 1622 cm⁻¹ (see Supporting Information) was found to be +0.02 Å. Such an IR excitation-induced elongation is expected to affect the H-bond strength and to reduce the DA coupling matrix element that determines the ET rate.⁷

While the lifetime of the excited bending modes is expected to be ca. 1 ps, other modes with lower frequencies will be populated via vibrational relaxation. There are ~10 other modes in DBA that will drive asymmetric H-bond length changes, allowing dynamical modulation of the donor—acceptor coupling. If these changes are fast on the ET time scale, the mean squared DA coupling will enter the ET rate constant.⁴ Another mechanism that could reduce the DA coupling matrix element is a dynamic reduction of the guanine and/or cytosine ring planarity, as deformation modes are excited by vibrational relaxation. Indeed, nucleic acid intramolecular distortions were indicated as controlling electronic coupling fluctuations in DNA and PNA ET.¹⁷ The planarity of the bridge hydrogen-bonded pair is expected to influence the DA coupling and the CS rate significantly.⁷,¹⁴,¹⁵ Modulating the planarity of the G and C rings may also enhance the nonradiative relaxation of the excited state to the ground state, reducing the quantum yield of the CS reaction.¹⁵ While the Watson—Crick structure is the dominant one, the presence of other tautomers in quantities up to 2% has been predicted, most notably the imino-enol double hydrogen atom transfer structure.¹⁵,¹⁶ The influence of IR on the tautomer distribution in the GC complex may affect the ET rate. It is also possible that inelastic ET channels may contribute to the observed effect.¹,³

In summary, we have demonstrated, for the first time, that DA charge transfer can be modulated by direct vibrational excitation of bridging modes. Dynamic modulation of the effective DA coupling, by partial disruption of the bridging H-bonds and/or by distortion of the planarity of the bridging π-electron system, is proposed to explain the effect. Several approaches can be envisioned to enhance the effect of modulation, including the use of DA pairs with higher symmetry or those with a better match between the lifetimes of the vibrational modes and the ET time scale. Ultimately, such improvements could lead to the development of new molecule-based charge separating devices, including molecular switches, wherein IR irradiation is used to control the underlying ET kinetics.

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Supporting Information Available: Complete ref 10, computational details, and extended FTIR spectrum. This material is available free of charge via the Internet at http://pubs.acs.org.

References