The Humoral Response to HIV-1: New Insights, Renewed Focus

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During the past 2 decades, significant advances in our understanding of the humoral immune response to human immunodeficiency virus type 1 (HIV-1) infection have been made, yet a tremendous amount of work lies ahead. Despite these advances, strategies to reliably induce antibodies that can control HIV-1 infection are still critically needed. However, recent advances in our understanding of the kinetics, specificity, and function of early humoral responses offer alternative new approaches to attain this goal. These results, along with the new broadly neutralizing antibody specificities, the role for other antibody functions, the increased understanding of HIV-1–induced changes to B cell biology, and results from the RV144 “Thai” trial showing potential modest sterilizing protection by nonneutralizing antibody responses, have renewed focus on the humoral system. In this review, recent advances in our understanding of the earliest humoral responses are discussed, highlighting presentations from the meeting on the Biology of Acute HIV Infection.

Since the discovery of human immunodeficiency virus type 1 (HIV-1), study of the antibody response to the virus has been integral to the search for mechanisms of virus inhibition and the development of strategies to prevent infection. The initial antibody response to HIV-1 is primarily directed at nonneutralizing epitopes on envelope glycoprotein (Env), whereas antibody responses that neutralize transmitted and early infection virus isolates are delayed, arising several months after the transmission event [1]. Broadly neutralizing antibody (bnAb) responses arise in a minority of patients and are generally not associated with control of viremia [2–4], and in most patients neutralizing antibody (nAb) responses lag behind virus escape [5, 6]. More than 2 decades of research has yielded a small number of human monoclonal antibodies capable of neutralizing a wide range of virus isolates [7–14] and many antibodies with narrow breadth [15]. Furthermore, changes in the humoral immune system have been described in patients with AIDS and include abnormal B cell activation [16], a loss of recall responses to antigens [17], and the development of autoantibodies and other autoimmune manifestations [18, 19]. Passive protection trials in non-human primates have shown that monoclonal antibodies of sufficient titer and quality can be protective against live virus challenge [20, 21], and many vaccine efforts have been aimed at inducing bnAbs. Unfortunately, no vaccine candidates have reliably induced antibodies similar to the rare human bnAbs [22, 23].

Several recent findings have reenergized interest in the potential of protective nAbs against HIV-1, either induced through vaccination or given in passive transfer. In addition to the well-characterized nAbs that bind to Env glycans (2G12 [7]), the Env gp41 membrane-proximal external region (MPER) (2F5 [8, 12], 4E10 [11], Z13 [9, 14]), and the Env gp120 CD4-binding site (immunoglobulin [Ig] G1b12 [10]), another target for bnAbs involving the Env gp120 V1/V2 and V3
regions, have been identified [13, 24, 25]. This quaternary epitope is expressed on neutralization-sensitive trimeric Env and can be partially blocked by soluble CD4 binding [13, 24]. Type-specific nAbs to this region have been reported [24, 25], and recently 2 new bnAbs against this epitope were identified using a high-throughput screen of >30,000 activated memory B cells from 1800 infected donors [13]. This additional nAb target, coupled with the observation that nAb breadth occurs more frequently than previously thought—up to 20% of subjects ultimately develop a degree of breadth [4]—provides encouragement that such bnAbs can arise naturally and therefore that the right vaccine might also induce an nAb response.

Another hopeful note has been struck by the results of the ALVAC prime-recombinant Env gp120 boost RV144 phase III “Thai” trial [26]. In the modified intention-to-treat analysis, 31% vaccine efficacy was found, with suggestive evidence that protection occurred early after vaccination. Thus, the vaccine produced a weakly protective effect against acquisition but not viral control after infection, potentially related to the levels of binding antibodies elicited. Although the magnitude of the effect was small, these results are in stark contrast to the failures of other high-profile studies, including the VaxGen/AIDSVAX trial [27], the Merck Step trial [28], and the failed cellulose sulfate microbicide trial [29]. These new study results suggest that the kinds of protective responses desired from vaccines may be achievable.

On 22–23 September 2009, a symposium was held in Boston, Massachusetts, on the biology of acute HIV-1 infection. The humoral session of the Boston meeting reported on a wide range of aspects of the role of the humoral immune system in acute HIV-1 infection. The talks spanned topics including B cell development and germinal center formation, the evolution of the autologous and bnAb responses, and additional responses mediated by antibodies that may play a key role in containment of viral replication. These topics and an overview of related literature constitute the remainder of this review.

**B CELL DEVELOPMENT AND DYSFUNCTION**

The development of B cells occurs in the bone marrow with committed lymphocyte precursors that undergo sequential immunoglobulin gene rearrangements [30]. These rearrangements ultimately lead to the expression of surface immunoglobulins with specificity identical to that of the antibody secreted by the same cell [31]. B cells undergo initial positive and negative selection events before transitional 2 B cells emigrate from the bone marrow and become naive B cells [32]. Once naive B cells encounter antigen combined with support signals from T cells, they mature and become memory and plasma cells, which are responsible, respectively, for recall responses and for secreting high levels of plasma antibodies.

Unlike their T cell counterparts, antibody-secreting cells do not need to circulate in blood to perform their “effector” function. B cells in various maturational stages can circulate in the blood but also can reside in tissues, including specialized lymphoid tissues, such as the spleen, lymph nodes, tonsils, and bone marrow. Similar to T cells, B cells reside in other tissues and comprise a portion of the lymphocytes resident in mucosa-associated lymphoid tissues in the genitourinary and gastrointestinal tracts. As with T cells, it is thought that tissue B cells play a critical role in host defense from pathogens and commensal organisms. A number of studies of HIV-1–lymphoid tissues noted structural changes to B cell follicles, including follicular hyperplasia that correlated with the development of clinical lymphadenopathy [33, 34]. These early studies did not demonstrate destruction of B cell follicles until late in clinical disease [33, 34].

More recent studies have established that gastrointestinal tract CD4+ T cells are a principal site for HIV-1 and simian immunodeficiency virus replication [35, 36], that depletion of those cells occurs early during infection [37, 38], and that those T cells are not rapidly replaced after antiretroviral therapy is started [39]. These data, combined with those presented at the Boston meeting, suggest that HIV-1–associated damage to lymphoid tissues in the gastrointestinal tract, along with loss of CD4+ T cell help, may explain the delay in nAb production after HIV-1 transmission [40]. This study focused on the effect of HIV-1 infection on the humoral immune compartment in patients with acute or early HIV-1 infection, identified through clinical protocols established by the Center for HIV/AIDS Vaccine Immunology. Patients donated peripheral blood and/or terminal ileal biopsy specimens that were analyzed by means of immunohistochemistry and quantitative image analysis as well as flow cytometry. Using a variety of markers to identify both primary and secondary germinal centers in terminal ileal tissues, the study found evidence of germinal center damage in >80% of germinal centers identified in specimens from patients with acute or early HIV-1 infection, compared with <5% from healthy control tissues. Similarly, when germinal centers were analyzed for the presence of secondary follicle formation, indicative of B cell selection and maturation with support from follicular dendritic cells and T cells, two-thirds of follicles studied from healthy tissues showed evidence of secondary germinal center formation, compared with one-third of those from HIV-1–infected patients. Together, these data showed that, in terminal ileum, the environments that support B cell maturation were severely damaged in HIV-1 infection. These data contrasted with those previously described for other lymphoid tissues but correlated well with data on early T cell loss after HIV-1 transmission [35–39].

Flow cytometric analysis showed that in terminal ileum and peripheral blood there was a decreased fraction of B cells with a naive phenotype and a corresponding increase in B cells with
a memory B cell and plasma cell phenotype [40]. In particular, the fraction of circulating B cells with a plasmablast or plasma cell phenotype was higher in patients with acute or early HIV-1 infection [40], in some patients higher than the fraction reported for other models of antigen stimulation, such as influenza immunization [41]. The reason for this increase in circulating plasma cells in acute or early HIV-1 infection is not known, but it may be due to the high levels of cytokines circulating during early infection [42]. At this time, work is ongoing to determine the significance of the plasmacytosis observed during acute or early HIV-1 infection and to compare it with that seen in other disease and vaccination models.

**bnAb RESPONSES**

nAbs can be detected in most patients who are infected with HIV-1 at some point in the first year of infection after seroconversion [43]. This antibody response has been described as ineffective, because autologous nAbs often lag behind changes in the virus and rarely target the contemporaneous virus [6]. Alternatively, nAbs have been shown to drive virus escape [44] and may therefore be responsible for control of viremia in some patients after cessation of antiretroviral therapy [45]. nAbs can fall into 2 categories: antibodies that bind to variable regions of HIV-1 and that neutralize a relatively narrow group of isolates and those that bind to conserved regions of the virus and that can neutralize a wider range of isolates, including those in different subtypes or from diverse geographical regions [23]. For reasons that are not entirely clear, the initial antibody response to transmitted HIV-1 in most individuals is primarily narrow in its spectrum, with bnAbs produced in only a fraction of infected individuals, later in HIV-1 infection [4].

Recently, a number of new studies describing escape of HIV-1 from nAb pressure have begun to shed light on this phenomenon. All HIV-1–infected individuals produce autologous nAb responses that the virus rapidly evades [5, 6]. In her presentation, Morris postulated that identifying nAb specificities that evolve naturally may identify vulnerable regions of HIV-1 that the immune system is able to target. Given the difficulty in trying to elicit bnAbs thus far [22, 23], induction of these early nAb responses could serve as a strategy to induce protective nAbs through vaccination. To define the pattern and evolutionary kinetics of these autologous humoral response, autologous nAb activity was examined in a well-characterized acute infection cohort, the CAPRISA cohort [2, 46]. Although the kinetics of the autologous nAb responses varied, all subjects developed autologous nAbs, with a delay ranging from 9 weeks after infection up to 1 year, but these kinetics did not correlate with the rate of disease progression [2]. Autologous nAbs targeted particular regions of HIV-1 Env sequentially, with the C3 domain being the earliest target, followed by the V1/V2 region, and later other regions of the Env complex [47]. The nAb targets were confirmed after back-mutation of later viral variants, which were engineered to encode for earlier wild-type sequences that restored nAb sensitivity. In a representative example, 2 waves of nAbs were observed, and back-mutation of the later viral constructs demonstrated that these waves represented 2 different nAb specificities that developed at distinct time points after infection, first to the C3 domain and then later to the V1/V2 regions of the viral Env. This staged response developed despite the fact that the second nAb epitope was present in the earliest viral isolates, suggesting that immunodominant patterns may drive reproducible antibody targeting of vulnerable regions of the viral Env [47]. In the other individuals studied, the pattern varied in such a way that although the nAbs targeted the predicted areas, escape mutations that conferred escape from nAb recognition occurred at a distance from these regions of the viral Env.

The phenomenon of sequential antibody responses to transmitted HIV-1 has been described in other contexts (Figure 1). The earliest detectable humoral responses to transmitted HIV-1 can be detected in plasma donor pools and were described by Tomaras et al [1]. These responses consist of immune complexes of antibody and HIV-1 virions that arise on average 8 days after the onset of viremia, followed in 5 days (on average 13 days after the onset of viremia) by free antibodies directed against Env gp41. In a majority of samples studied, the initial antibody response consisted of simultaneous IgM and IgG antibodies, suggesting early class switching associated with HIV-1 infection. Interestingly, compared with Env gp41 antibodies, gp120 antibody responses were delayed, occurring on average 28 days after the onset of viremia; this phenomenon occurs despite simultaneous exposure of the immune system to both antigens. Although waves of antibody responses to different viral antigens are not unknown (eg, patterns of antibodies associated with Epstein-Barr virus infections [48]), these antigens are often involved in different stages of the virus life cycle. The reasons for the development of sequential antibody responses to HIV-1 Env are as yet unknown.

Additional data presented by Morris showed that in some patients, broad autologous nAb responses may occur at early time points. One particular subject, CAP206, developed an anti-MPER antibody detectable after only 6 months of infection [49]. This nAb was able to inhibit a wide range of viruses, and the activity was absorbable by a peptide with the MPER sequence. Using a number of techniques, this group of investigators is pursuing further studies to isolate the antibody and determine whether it was a novel B cell clone that arose spontaneously or the result of affinity maturation of an original autologous nAb response.

To determine the impact of autologous nAbs on antiviral control and viral evolution, this group used quantitative polymerase chain reaction to track fluctuations of wild-type and
emerging quasispecies or viral variants [47]. This group was able to demonstrate that after the appearance of the first nAb wave, the overall viral load declined consistent with the decline in the wild-type viral variant levels. This was followed by the emergence of an escape variant that then outcompeted the wild-type isolate, resulting in a slight increase in the overall viral load. This observed “blip” in overall viral load in parallel with antibody-induced emergence of alternate viral quasispecies suggests that similar fluctuations in viral load may reflect the emergence and disappearance of viral variants under immune pressure.

Another set of studies focused on HIV-1 subtype C was presented by Derdeyn of Emory University. Serodiscordant couples from a cohort established in Lusaka, Zambia, were recruited and the HIV-1–negative partner was tested every 3 months until infection occurred; after infection, the partners provided longitudinal samples every 3 months [50]. Previous work from this cohort demonstrated that autologous nAbs in some patients were detectable at 2 months after infection, suggesting that the antibody response in this group of subtype C infected individuals might differ from that found in subtype C infection in the CAPRISA cohort or in subtype B infection [51]. The majority of subtype C infections in this cohort appeared to be from a single transmitted virus [52], consistent with similar findings in subtype B infection [53]. Using a single-genome amplification technique, the group cloned functional env genes and generated unmutated pseudoviruses as well as chimeras and mutants that were used in a single-round neutralization assay to study escape [50]. Consistent with reports from other investigators, virus isolates resistant to neutralization with serum samples at a given time could be detected at every time tested, suggesting that the antibody response in these patients lagged behind HIV-1 mutations.

Using chimeric and mutated Env, a number of interesting findings emerged. No single Env domain or restricted set of Env domains was found to dominate the autologous nAb response. In contrast to subtype B infections, the V3 loop was not a prominent epitope for nAbs, but, similar to data from the CAPRISA cohort, there was evidence of escape mutations in V1/V2, V5, the α2 helix, and the gp41 ectodomain. In 1 patient, changes in glycosylation were found to be associated with escape, both with glycan loss in V1 and with glycan gain in V2. Most intriguingly, a pattern of convergent pathways was found for 1 patient at 28 months after infection; for 1 Env isolate, changes to V5 conferred resistance, but for a different Env isolate, resistance was conferred by changes to V1/V2 and...
the gp41 ectodomain. Thus, similar to observations by the Morris group, these data from the Zambia cohort confirm that the nAb response may not always drive HIV-1 mutation in a linear fashion but that it may select different mutational profiles through sequence, glycosylation, or structural features of the Env, depending on other features of a given viral quasispecies. These data suggest that although predictable patterns exist in the kinetics of protein targeting in acute HIV infection, the pathways of escape may be more complicated.

**ANTIBODIES TO RECRUIT INNATE IMMUNITY**

Antibody functions not directly related to their ability to neutralize HIV-1 can play a critical role in prevention of infection. In fact, the constant region of the antibody is able to recruit a number of additional effector functions, including the deposition of the complement system, stimulation of cytokine or chemokine secretion, triggering of phagocytosis, immunoregulation, and recruitment of cytolytic activity [54]. All of these activities are triggered after the interaction of the CH2 domain of the antibody with circulating complement or Fc receptors found on immune cells (Figure 2) [55, 56]. Given that Fc receptors are present on all innate immune cells (including natural killer [NK] cells, dendritic cells, and monocytes) as well as on B cells, antibodies also play a central role in orchestrating and providing specificity for the potent antiviral activity of diverse innate immune cells. The roles of Fc receptor binding and complement fixation are not equally involved in antibody-mediated protection of simian immunodeficiency virus infection, as demonstrated by modifications of nAb IgG1b12 [57]. Sterilizing protection from infection was unaltered by changes to the Fc region of IgG1b12 that abrogated complement recruitment, whereas modifications that resulted in a loss of Fc receptor engagement decreased the protective antiviral nAb potency, suggesting that the ability of this nAb to recruit the cells of the innate immune system via Fc receptors may play a critical role in antiviral control [57]. Similarly, the efficacy of some therapeutic antibodies, such as the anti-CD20 antibody rituximab used for the treatment of B cell diseases such as non-Hodgkin lymphoma and Epstein-Barr virus–driven posttransplant lymphoproliferative disorders, hinge on the capacity of that antibody to recruit the effector functions of innate immune cells, such as NK cells [58, 59]. Several studies have shown that antibody-dependent cell-mediated cytotoxicity (ADCC)–inducing antibodies can play a dramatic role in the early containment of several infections, but less is known about the role of this humoral immune function in the early control of HIV-1 infection [56].

To define the role of ADCC-inducing antibodies in antiviral control, the ADCC activity of antibodies was compared among HIV-1–infected individuals using the antibody-dependent cell-mediated virus inhibition assay described by Forthal et al [60, 61]. This activity was observed only in a fraction of chronically HIV-1–infected patients who had active viral replication, whereas strong activity was observed in individuals with undetectable viral loads associated with either antiretroviral therapy or spontaneous elite control. An assay based on multiparameter flow cytometry showed that antibodies from elite controllers were also able to recruit NK cell degranulation more effectively and with a broader polyfunctional profile of NK cell responses compared with antibodies from chronically infected individuals. Differences in the capacity of antibodies to recruit NK cell activity probably pertain to intrinsic differences in the antibodies generated in different patient populations, given that effector (NK) cells and targets are all derived from HIV-negative control subjects. These results strongly suggest that, despite the fact that elite controllers typically exhibit reduced levels of nAbs [62], these individuals do elicit antibodies with properties that enable them to recruit the antiviral activity of NK cells, potentially contributing to their antiviral control.

In contrast to the elite controllers, only a fraction of acutely HIV-1–infected individuals exhibited ADCC–inducing antibodies at baseline; however, in those individuals who exhibited ADCC activity, the activity was as potent as the antiviral activity of antibodies from elite controllers. Furthermore, ADCC-inducing antibodies from acutely infected individuals were also able to elicit polyfunctional NK cell responses. These data, combined with the fact that antiretroviral therapy was able to partially reconstitute ADCC activity, strongly suggest that persistent viral replication—rather than high antigen loads, as seen in acute HIV infection—may be responsible for a loss of
ADCC-inducing activity in chronic progressive HIV-1 infection.

In addition to these changes in the quality of antibody-mediated recruitment of innate immune activity, effective ADCC is contingent on the presence of a functional effector cell bearing an Fc receptor or receptors [56]. Previous work has shown that ADCC activity declines with progressive infection, and though the antibody response has a diminished capacity to inhibit viral replication, it is also plausible that waning ADCC activity may be attributable to changes in Fc receptor expression in cells over the course of HIV-1 infection. In fact, defining the pattern of Fc receptor changes that occur throughout HIV-1 infection may provide us with clues regarding the mechanism by which innate immune recruiting antibodies may mediate their antiviral control. Furthermore, it is possible that ADCC-inducing antibodies may be particularly important during acute infection when viral loads decline dramatically after peak viremia [63]. Indeed, Fc receptors were dramatically altered on the surface of innate immune cells at different stages of HIV infection, with a reduction in Fc receptor expression on innate immune cells in chronic, progressive HIV-1 infection. In contrast, Fcy receptor 1 expression was significantly enhanced on monocytes in acute HIV-1 infection. However it is still unknown whether this elevation is up-regulated after all viral infections or only in response to HIV-1 specifically or whether innate immune recruiting capacity is involved in the clearance and control of the virus early in infection.

CONCLUSION

Over 2 decades, significant advances in our understanding of the humoral immune response to HIV-1 infection have been made, yet a tremendous amount of work lies ahead. Despite these advances, we are still working to create strategies to reliably induce antibodies that can control HIV-1 infection. Antibodies against HIV-1 can display a wide array of reactivity including rare bnAbs against gp41 MPER, potent CD4-binding site antibodies, and antibodies against the glycan shield and other conserved epitopes. These types of antibodies are, of course, the primary targets of the humoral immune response to HIV-1 infection. However, to date these antibodies have not been found early in infection nor have they been readily elicited by HIV-1 vaccine candidates. In contrast, antibodies that are commonly elicited by both vaccination and infection have not been considered capable of mediating protection, including weakly neutralizing CD4 antibodies, coreceptor binding site and hypervariable region antibodies, and gp41 non-neutralizing antibodies.

Recent developments in our understanding of autologous neutralizing antibodies as well as Fc receptor-mediated anti–HIV-1 activity offer new approaches to protection from HIV-1 transmission at the mucosal surface. Moreover, data from the RV144 Thai trial suggest a modest level of sterilizing protection likely to have been elicited by nonneutralizing antibodies, raising the hypothesis that potentially narrower breadth and/or innate immune recruiting capacities may have contributed to preventing acquisition. Intense efforts are underway to dissect both the cellular and humoral immune responses in this vaccine trial to define potential correlates of protection. These results, along with the new bnAb specificities, the potential role for other antibody functions, and the increased understanding of HIV-1–induced changes to B cell biology, have renewed focus on the humoral system and efforts to recruit the system as a whole rather than just its parts.

References