

Testosterone secretion varies in a sex- and stage-specific manner: Insights on the regulation of competitive traits from a sex-role reversed species

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ABSTRACT

Testosterone (T) mediates a variety of traits that function in competition for mates, including territorial aggression, ornaments, armaments, and gametogenesis. The link between T and mating competition has been studied mainly in males, but females also face selection pressures to compete for mates. Sex-role reversed species, in which females are the more competitive sex, provide a unique perspective on the role of T in promoting competitive traits. Here, we examine patterns of T secretion in sex-role reversed northern jacanas (*Jacana spinosa*) during breeding, when females are fertile and males are either seeking copulations or conducting parental care. We measured baseline levels of T in circulation along with a suite of behavioral and morphological traits putatively involved in mating competition. We evaluated hypotheses that levels of T track gonadal sex and parental role, and we begin to investigate whether T and competitive traits co-vary in a sex- and stage-specific manner. Although females had higher expression of competitive traits than males at either breeding stage, we found that females and incubating males had similar levels of T secretion, which were lower than those observed in copulating males. T was correlated with wing spur length in females and testes mass in copulating males, but was otherwise uncorrelated with other competitive traits. These findings suggest that levels of T in circulation alone do not predict variation in competitive traits across levels of analysis, including gonadal sex and parental role. Instead, our findings coupled with prior research indicate that selection for female mating competition and male care may generate different physiological regulation of competitive traits.

1. Introduction

For males of many vertebrate species, testosterone (T) has been associated with phenotypic traits that lead to successful competition over mates. These traits include aggressive behavior (Wingfield et al., 1990), weaponry (Malo et al., 2009), body size (Cox et al., 2009), ornamentation (McGlothlin et al., 2008), and gonadal size (Preston et al., 2012). Females also express many of these same traits, which likewise function in competition for mates and other breeding resources (Clutton-Brock, 2009; Hare and Simmons, 2018; Rosvall, 2013a; Tobias et al., 2012). Like males, females secrete T and have the physiological capabilities to respond to T (Staub and De Beer, 1997). However, the relationship between T and mating competition in females has found more equivocal support (Cain and Ketterson, 2012; Goymann and Wingfield, 2014; Rosvall et al., in press), suggesting that T may influence trait expression differently in males and females. This can be explained in part by differences in the selective pressures that drive

endocrine mechanisms of behavior (Wingfield et al., 1990). For instance, the relative importance of aggression versus parental care in males and females may shape both sexual dimorphism in T, as well as behavioral sensitivity to T (Lynn, 2008; Rosvall, 2013b) – this is one hypothesized driver for why females tend to have lower levels of T in circulation than males.

Levels of T in circulation are typically elevated at the beginning of the breeding season, when competition for mating opportunities is high, and then T levels decline as behavioral efforts shift to parental care (Wingfield et al., 1990). This cross-stage shift in T production has been well demonstrated in males (Hirschenhauser and Oliveira, 2006), and is potentially more dramatic in females (DeVries et al., 2012; George and Rosvall, 2018; Jawor et al., 2007). Cross-stage shifts may also alter correlations between T and sexually selected traits, such that these traits are strongly integrated with T during competition for mates, but more independent from T during periods of parental care (Ketterson et al., 2009; Lipshutz et al., 2019a).

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In sex-role reversed species, females face stronger selection to compete for mates, and males predominantly care for offspring (Emlen and Oring, 1977), providing an opportunity to disentangle parental role from gonadal sex as drivers of variation in T. Whereas sex-role reversed females are freed from the constraints of parental care, selective pressures to care for offspring may limit T levels in males. The hypothesis that sex-role reversed females have higher T than males has intrigued behavioral ecologists for decades. Whereas early studies in sex-role reversed phalaropes found higher T in ovarian than testicular tissue (Höhn, 1970; Höhn and Cheng, 1967), most studies have found that the direction and magnitude of sex differences in T is typically similar between role-reversed species and species with traditional sex roles (Fivizzani et al., 1986; Gratto-Trevor et al., 1990; Rissman and Wingfield, 1984; Goymann et al., 2004; but see Muck and Goymann (2011)). Approaching sex-role reversed species from a hormonal perspective can shed light on the regulation of competitive traits, which may vary based on gonadal sex and/or parental role.

One of the most well-known examples of sex-role reversal is the jacana (family Jacanidae). Although both sexes aggressively compete for territories, females face more intense sexual selection, have higher potential reproductive rates, and have female-biased operational sex ratios (Emlen and Wrege, 2004a). Female jacanas also have larger secondary sexual traits, including weaponry, body mass, ornamentation, and behavioral dominance (Emlen and Wrege, 2004a; Lipshutz, 2017; Stephens, 1984). Males provide nearly all parental care, which includes incubation and foraging with chicks (Emlen and Wrege, 2004b; Jenni and Collier, 1972). Although jacanas are a classic example of female-biased dimorphism in behavior and morphology (Emlen and Oring, 1977), there is currently no published work on the physiological mechanisms that regulate the expression of these traits, limiting our ability to connect proximate mechanisms with behavioral and evolutionary processes in this well studied system.

Here, we quantified circulating T levels in free-living, sex-role reversed jacanas to evaluate the hypotheses that levels of T track gonadal sex and/or parental role. Sex-role reversed females should be unconstrained by parental care, and if they have higher T than males, this would suggest that T levels are shaped by parental roles more so than gonadal sex. We further predict that males incubating eggs should have lower T than males seeking copulations. As a secondary goal, we begin to evaluate whether co-variation between T and competitive traits varies in a sex- and breeding stage-specific manner. Of the studies on sex-role reversed species that examined phenotypic co-variation with T, one reported that T positively correlated with plumage coloration and body condition in females only (Muck and Goymann, 2011), and another found that ornaments were testosterone-dependent in both sexes (Eens et al., 2000). Neither of these studies examined the influence of breeding stage on phenotypic co-variation, which is important for considering how temporally variable selection pressures (i.e. mating competition vs. parental care) shape differences both between and within the sexes. We predict that T will be more strongly linked with competitive traits in females and copulating males, but that T should be more independent from these traits in incubating males.

2. Materials and methods

2.1. Subjects

Northern jacanas (*Jacana spinosa*) are tropical shorebirds found throughout Central America, from Mexico to Panama. They breed asynchronously and year-round, although breeding increases during the rainy season in Panama from roughly May to October. We conducted fieldwork from 4 June to 9 July 2018 in La Barqueta, Chiriqui, Panama (8.207N, 82.579W).

2.2. Aggression assays

Our first step was to measure aggression, a behavioral component of the competitive phenotype. We first observed each individual for several days to ensure that it was pair-bonded (e.g. foraging with mate) and territorial (e.g. actively defending territory from intruding floaters). Territorial residents behave distinctly from floaters, who do not breed nor defend territories (Emlen and Wrege, 2004a). We also determined breeding status based on whether males were copulating ($n = 5$) or incubating a nest ($n = 7$). We did not color-band individuals as in previous studies (Lipshutz et al., 2019b) and were therefore unable to determine harem size, but territory holders have high site fidelity (Emlen and Wrege, 2004a), and the agricultural land where we conducted this research has small ponds and canals where territorial boundaries are distinct. We only sampled individuals that we observed in these same stable locations consistently each and every day.

We assayed aggression in males ($n = 12$) and females ($n = 10$) using a random combination of 4 taxidermic female mounts and 4 conspecific vocalizations following Lipshutz (2017). Briefly, we set up a camouflage blind and placed the mount and speaker in the center of a female and one of her male mates' territory (~15 m from the nest if the male was incubating). We used a 10-s recording of jacanas fighting and vocalizing to attract the focal individual, and began the 5-min assay the moment the female or male responded. We measured a suite of aggressive behaviors, including average distance from the mount, hover flights, wing spreads, flyovers, and vocalizations, described previously in an ethogram for jacanas (Lipshutz, 2017). We tested females and males independently of each other, except for one pair.

2.3. Capture and plasma collection

Next, we sought to measure baseline plasma T levels in these same individuals. T levels can change in response to social stimulation, but most studies suggest this process takes time (Gleason et al., 2009; Hirschenhauser et al., 2003; Oliveira et al. 2002), consistent with time-course studies in birds showing that T levels peak about 30 min after activation of the HPG axis (Jawor, 2006; Rosvall et al., 2016). Therefore, we attempted to capture each bird immediately after the short 5-min aggression assay and measure T levels before they could be affected by this simulated intrusion. We collected individuals using an air rifle, followed by an anaesthetic overdose of isoflurane and decapitation to collect trunk blood; brain and other tissues were reserved for future study.

We were able to collect 9 individuals quickly after the intrusion (average time from intrusion start to euthanasia = 9 min 20 s \pm 1 min 35 s). We were unable to collect 13 individuals within ~10 min post-intrusion, and so, we returned 5–8 days later to collect these individuals ($n = 6$ females, 6 males). For these 'delayed' collections, we ensured that male breeding stage did not change; for instance, if a male was incubating on the day of the aggression assay, we monitored him daily with behavioral observations to confirm that he was still incubating on the day of collection. For each sex, we balanced immediate and delayed collection sample sizes. In addition, we tested for and found no difference in T levels between immediate vs. delayed samples for either females ($t = 0.51$, $df = 7.42$, $p = 0.62$) or males ($t = -0.24$, $df = 9.84$, $p = 0.81$), so we combined these immediate vs. delayed collection individuals for further analysis. These findings are consistent with recent analyses that simulated territorial intrusions do not lead to rapid increases in T secretion in birds (Goymann et al., 2019; Wingfield et al., 2019) but see Wingfield and Wada (1989).

We collected whole blood into heparinized BD Microtainers (product #365965) and stored on an ice pack for up to 6 h until we separated plasma by centrifuging for 10 min at 10,000 rpm. We stored plasma at -20 °C for later testosterone assays.

2.4. Morphological measurements

Postmortem, we measured several traits putatively involved in mating competition in jacanas: wing spur length, body mass, facial shield length, and gonad mass. In a congener, the wattled jacana (*J. jacana*), territorial resident status was associated with larger wing spurs, facial shields, and body mass for both sexes, and only territory holders can breed (Emlen and Wrege, 2004a). These traits are much smaller in floaters, who do not obtain territories or reproduce. We assume that these traits similarly relate to intra-sexual competition in northern jacanas, although we have not tested this explicitly. For both sexes, wing spurs, facial shields, and body mass are significantly larger in adult female jacanas compared to males (Emlen and Wrege, 2004a; Lipshutz, 2017), but these traits are small in juveniles of both sexes (Lipshutz, personal observation).

We measured body mass with a digital scale (0.01 g), and we measured wing spur length (from base center to tip, 0.1 mm) with calipers. Wing spurs are sharp, yellow keratinous sheaths over metacarpal bone growths that jacanas use as weapons and display during aggressive posturing. In the wattled jacana, wing spurs length positively correlated with age in males but not females, perhaps because they were worn down by abrasion from fighting (Emlen and Wrege, 2004a). However, we did not observe any worn down wing spurs in this study. In northern jacanas, facial shields are yellow, fleshy, and extend from the upper mandible to the forehead. We measured facial shield length (from right nare to top right lobe, 0.1 mm) with calipers. We also measured gonad mass with a digital gem scale (0.1 oz). Gonad mass has clear connections to mating competition because jacanas are polyandrous – males that are simultaneously mated with a single female compete to fertilize her eggs, and females are continuously producing eggs for available mates to incubate (Emlen et al., 1989). Testes size is associated with T in many avian species (Garamszegi et al., 2005), and testes size is correlated with sperm length in shorebirds (Johnson and Briskie, 1999). Visual inspection of the gonad(s) was also used to confirm sex and breeding stage. All 7 males that we had observed incubating had brood patches, and one male we observed copulating had feathers growing in over his brood patch, suggesting he had recently terminated incubation. All 10 females were fertile, with hierarchical ovarian follicles.

2.5. Testosterone enzyme immunoassay

We extracted steroids from plasma samples using diethyl ether (3 × extractions) and reconstituted in 250 µL assay buffer. We measured testosterone using a High Sensitivity Testosterone ELISA kit (Enzo #ADI-900-176, Farmingdale, NY, USA) following methods described in George and Rosvall (2018). The kit reports a detection limit of 2.6 pg/mL. Adjusted for buffer reconstitution volume and plasma volume, this amounts to a detection limit of 0.0325 ng T/mL plasma in males and 0.01625 ng T/mL plasma in females. All of our sample values were above this detection limit. We confirmed assay parallelism by comparing concentrations from a standard curve made by kit standards to a displacement curve made from dilutions of a copulating male jacana's plasma ($R^2 = 96.3\%$). We ran all samples in duplicate. We initially used 40 µL plasma from females, and 20 µL from males. Samples from three females and four copulating males initially showed less than 20% maximum binding, and two males showed greater than 80% maximum binding, so we re-ran them using 10 µL plasma and 40 µL of plasma, respectively, to obtain values in the most sensitive part of the curve. We calculated T concentration by comparing sample absorbance with the absorbance of the assay's standard curve (Gen5 curve-fitting software, Biotek EPOCH plate reader, Winooski, VT, USA). Intra-assay CV was 4.44% and inter-plate CV was 5.94%.

Table 1

Loadings for principal component analysis of aggressive behavior.

Vocal Parameter	PC1
Eigenvalue	1.79
Proportion of variance	64.14%
Distance to mount	-0.41
Hover Flights	0.43
Wing Spreads	0.46
Flyovers	0.46
Vocalizations	0.47

2.6. Statistical analysis

We conducted all statistics in R version 3.6.1 (R-Core-Team, 2019). We examined normality using a Shapiro-Wilk normality test and examined outliers using a Grubbs test in the R package 'outliers' (Komsta, 2006). We normalized T using a log scale transformation for all statistical comparisons. To compare T, morphology, and aggression between the sexes at different male breeding stages, we used a one-way ANOVA, followed by a Tukey's post hoc test. Comparisons between sexes or male breeding stages were made using Student's *t* tests or Wilcoxon tests, depending on normality. To assess the degree of covariation among T and competitive traits, we used Spearman's correlations. To control for multiple testing, we used the Benjamini-Hochberg method with the *p.adjust* function in the R package 'stats' (R-Core-Team, 2019).

We also summarized the 5 aggressive behaviors with a principal component analysis (PCA) using the *prcomp* function. We retained 1 PC with an eigenvalue > 1 (hereafter 'Aggression PC1'), which explained 64.1% of the variation in aggressive behaviors (Table 1). Distance loaded negatively onto Aggression PC1, and hover flights, wing spreads, flyovers, and vocalizations loaded positively, such that a more positive PC1 reflects a more aggressive response.

3. Results

3.1. Circulating testosterone varies by sex and male breeding stage

When all males were combined, male and female T level were not significantly different ($t = -1.58$, $df = 17.47$, $p = 0.13$). When considering male breeding stage however, the groups were significantly different ($F_{2,19} = 8.2$, $p = 0.0027$; Fig. 1). Copulating males had significantly higher T levels than incubating males (5.26 ± 2.2 vs 0.54 ± 0.18 ng/mL, Tukey: $p = 0.005$). Females had T levels (0.51 ± 0.13 ng/mL) similar to incubating males and significantly lower than copulating males (Tukey: $p = 0.004$). One copulating male had T similar to the average T levels of incubating males, but was not an outlier among other copulating males (Grubbs' test; $p = 0.099$); upon collection he was observed to have new feathers growing in over his brood patch, suggesting recent cessation of incubation.

3.2. Competitive traits vary by sex and male breeding stage

We confirmed female-biased dimorphism in traits putatively involved in competition for mates, as has been previously demonstrated in jacanas (Emlen and Wrege, 2004a; Lipshutz, 2017). Females had significantly longer wing spurs (14.9 ± 0.3 vs. 10.2 ± 0.4 mm, $t = 10.23$, $df = 20.49$, $p < 0.0001$), larger body mass (155.9 ± 2.6 vs. 92.8 ± 1.7 g, $t = 21.89$, $df = 18.54$, $p < 0.0001$), and longer facial shields (28.8 ± 0.4 vs. 23.6 ± 0.4 mm, $t = 9.69$, $df = 19.0$, $p < 0.0001$) than males in both breeding stages. Between males in different breeding stages, incubating males had significantly lower testis mass than copulating males (0.010 ± 0.0021 vs. 0.020 ± 0.0028 oz, $t = -2.57$, $df = 8$, $p = 0.033$), and a trend

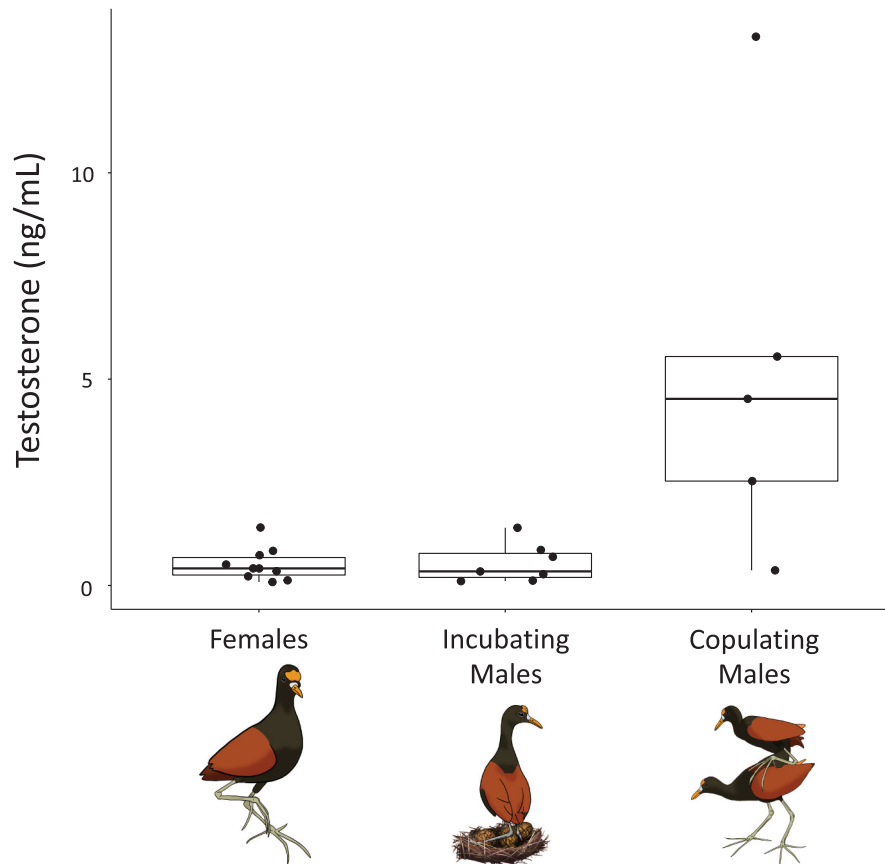


Fig. 1. Levels of testosterone in circulation in female (n = 10) and male jacanas at different breeding stages, incubating (n = 7) and copulating (n = 5). Boxplot horizontal lines represent median values. Illustrations by Mae Berlow.

towards longer wing spurs (10.8 ± 0.2 vs 9.4 ± 0.6 mm, $t = 2.08$, $df = 5.15$, $p = 0.091$).

Groups differed significantly in Aggression PC1 ($F_{2,19} = 3.89$, $p = 0.038$). Males defending their eggs (i.e. incubating males) were marginally more aggressive than copulating males (Tukey: $p = 0.089$) and significantly more aggressive than females (Tukey: $p = 0.049$). Aggression did not differ between females and copulating males (Tukey: $p = 0.99$).

3.3. Co-variation between testosterone and competitive traits

In females, T correlated positively with wing spur length, but not with other putatively competitive traits (Table 2). T was positively

Table 2
Spearman correlations (r_s) of circulating testosterone with competitive traits for females and males that were incubating or copulating. Significant relationships after Benjamini-Hochberg correction ($p \leq 0.05$) are bold.

	Females	Incubating Males	Copulating Males
Wing Spur Length	$r_s = \mathbf{0.83}$ $p = \mathbf{0.036}$	$r_s = -0.07$ $p = 0.91$	$r_s = -0.70$ $p = 0.58$
Body Mass	$r_s = 0.09$ $p = 0.91$	$r_s = 0.29$ $p = 0.71$	$r_s = 0.70$ $p = 0.58$
Facial Shield Length	$r_s = -0.21$ $p = 0.71$	$r_s = 0.57$ $p = 0.58$	$r_s = 0.4$ $p = 0.71$
Gonad Mass	$r_s = 0.25$ $p = 0.71$	$r_s = -0.64$ $p = 0.58$	$r_s = \mathbf{0.97}$ $p = \mathbf{0.036}$
Aggression PC1	$r_s = -0.061$ $p = 0.91$	$r_s = -0.29$ $p = 0.71$	$r_s = 0.45$ $p = 0.71$

correlated with testes mass among copulating males, but uncorrelated with any trait among incubating males. T was not correlated with Aggression PC1, regardless of sex or male breeding stage. Individual correlations for each group are plotted in a Supplemental figure (Fig. S1).

4. Discussion

In wild female and male jacanas, we found that differences in T secretion were contingent on male breeding stage, rather than sex alone. Circulating T levels in females were similar to levels observed in males conducting parental care. This is reflective of patterns found in other sex-role reversed species, for which females have similar levels of T in circulation to incubating males, despite female jacanas facing strong mating competition (Emlen and Wrege, 2004a). Copulating males had higher T than incubating males, suggesting that T and parental care may constrain one another. Although T varied by sex and male breeding stage in ways that suggest T is shaped by trade-offs between competition and care, we did not find widespread or consistent co-variation between individual differences in T and competitive traits.

In male jacanas, T differs by breeding stage, suggesting that levels of T are shaped by shifting selection pressures from mating effort to parental effort. This pattern is similar to studies of other sex-role reversed species, in which T was similarly low for both females and nesting males, and higher in courting males (Table 3). Stage-related variation in T is also prevalent in non-sex-role reversed species with male parental care (Wingfield et al., 1990). Experimental evidence that T reduces male parental care in role reversed spotted sandpipers (*Actitis macularia*) (Oring et al., 1989) as well as socially monogamous songbirds (Goymann and Davila, 2017; Ketterson et al., 1992; Van Roo, 2004; but see Lynn (2008)) indicates that high T can inhibit male parental

Table 3Levels of testosterone in circulation (Mean \pm SE) among sex-role reversed females and males in parenting and courting breeding stages.

Species	Females	Parenting Males	Courting Males	Citation
African Black Coucal <i>Centropus grillii</i>	0.57 \pm 0.06	0.59 \pm 0.14	2.16 \pm 0.51	Goymann et al. (2004)
Wilson's Phalarope <i>Phalaropus tricolor</i>	0.51 \pm 0.1	0.63 \pm 0.15	3.61 \pm 1.12	Fivizzani et al. (1986)
Red-necked Phalarope <i>Phalaropus lobatus</i>	0.085 \pm 0.047	0.22 \pm 0.055	3.98 \pm 0.73	Gratto-Trevor et al. (1990)
Spotted Sandpiper <i>Actitis macularius</i>	0.18 \pm 0.03	0.17 \pm 0.05	0.95 \pm 0.41	Rissman and Wingfield (1984)
Barred Buttonquail <i>Turnix suscitator</i>	0.2 \pm 0.1	NA	0.5 \pm 0.1	Voigt (2016)
Northern Jacana <i>Jacana spinosa</i>	0.51 \pm 0.13	0.54 \pm 0.18	5.26 \pm 2.2	this study

behavior, regardless of whether males are the more competitive sex. This idea was supported by a meta-analysis across vertebrates, finding that male parental care predicted T but mating system did not (Hirschenhauser and Oliveira, 2006), and another meta-analysis in humans, finding that fathers have lower T than men without children (Grebe et al., 2019).

Given the constraints of egg production and parental care typically imposed on females, T levels ought to be less correlated with suites of integrated traits in females than in males (Ketterson et al., 2009). However, we did not find strong phenotypic co-variation with T in either sex, except for wing spur length in females and testes mass in copulating males. Considering limited sample sizes and the potential for type II error, we view these analyses as a preliminary but important step towards understanding hormonal regulation of competitive traits and how it may vary based on gonadal sex and parental role. Females with higher T have longer wing spurs, a weapon used to fight over territories and mates. A similar finding in sex-role reversed barred buttonquail females (*Turnix suscitator*) indicated that female but not male levels of T positively correlated with body condition and the size and blackness of the melanin throat patch (Muck and Goymann, 2011). In moorhens, (*Gallinula chloropus*), the heaviest females, which tend to win most of the competitive interactions, also had higher T levels than lighter females (Eens and Pinxten, 2000). Indeed, female jacanas generally have longer wing spurs than males, but do not have higher T than males. In copulating males, only testes mass was correlated with T, a pattern that likely relates to spermatogenesis, and is supported in males of many avian species (Garamszegi et al., 2005). T did not correlate with aggression for either sex, regardless of male breeding stage. This is not unexpected, given that aggression and baseline levels of T do not correlate for many species (Kempnaers et al., 2008; Williams, 2008), although it is possible that such correlations would emerge when T secretion is at its physiological maximum (e.g. endogenous or exogenous activation of the HPG axis). This is an important avenue for future research, particularly considering how prior research on social responsiveness of T is biased towards temperate songbirds with conventional sex roles (Goymann et al., 2019). Our observation that incubating males had the lowest T and highest aggression suggests that mechanisms beyond T should be explored in the future (see below). Related to this point, higher aggression in incubating males may serve a parental purpose (i.e. defense of developing young, rather than territoriality or mating competition), and there is some evidence that different types of aggression may be regulated by different mechanisms (Duque-Wilckens and Trainor, 2017; Wingfield et al., 2006). Despite these complexities, our analyses nevertheless suggest sex- and stage-specific variation between T and competitive traits.

Our measurements of T were necessary to begin establishing the jacana system as a model for examining the role of gonadal steroids in orchestrating reproductive and parental behaviors. Circulating T did not sufficiently explain sex-role reversal; female jacanas express an exaggerated suite of competitive traits, but have low T levels on par with incubating males. An exciting next step is to explore how gonadal sex and parental role influence tissue-level regulation of competitive phenotypes. Beyond circulating T, other components of the androgenic signaling system could explain variation in competitive phenotypes, including variation in T production, metabolism, and/or sensitivity

(Ball and Balthazart, 2019; Fuxjager and Schuppe, 2018; Schmidt et al., 2008; Soma, 2006; Staub and De Beer, 1997). Tissue-specific variability in these components may regulate traits independently of circulating sex steroids (Bentz et al., 2019; Horton et al., 2014; Lipshutz et al., 2019a; Rosvall et al., 2012), and there is some evidence that tissue-specific regulation may be more prevalent for groups in which T levels are depressed (Demas et al., 2007; Rosvall, 2013a). Sex differences in androgenic signaling in the brain have also been found in several sex-role reversed species; female black coucals and barred button quails had higher mRNA expression of androgen receptors (AR) in neural regions implicated in the control of aggressive and sexual behavior (Voigt, 2016; Voigt and Goymann, 2007). Other hormones like progesterone could also explain variation in aggression, as was found in role-reversed black coucals (Goymann et al., 2008). In the future, we look forward to integrating proximate mechanisms more deeply with behavioral, morphological, and life history traits in jacanas, to better understand the physiological drivers of sex-role reversal.

As jacanas are tropical birds that can breed during all months of the year (Emlen and Wrege, 2004a), females may experience year-round selection in relation to mating competition. Territorial females breed simultaneously with multiple males in their harems, and males copulate and incubate asynchronously (Emlen and Wrege, 2004b). Although we did not follow females year round to assess individual fertility, all females in this study displayed clear signs of fertility (i.e. hierarchical ovarian follicles) regardless of whether or not their mates had nests. In contrast, males had large testes when copulating, but small testes when incubating. We hypothesize that female jacanas may have a continuously fertile gonadal state, similar to males of other species that maintain large, fertile testes throughout female incubation. Likewise, the cycling gonadal size of male jacanas from copulation to incubation is analogous to other species for which female ovaries return to a non-fertile state during incubation (Williams, 2012). The essentially continual state of female jacana fertility, paired with sex-role reversal, should generate stronger or more uniform selection on competitive traits, which are advantageous during most of the year and do not experience counter-selection in relation to wintering or migratory states. Our examination of T and competitive traits in jacanas add to a rich history of understanding how gonadal sex and parental roles influence mechanisms of behavior in sex-role reversed systems.

CRediT authorship contribution statement

Sara E. Lipshutz: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Writing - original draft, Writing - review & editing. **Kimberly A. Rosvall:** Funding acquisition, Supervision, Resources, Writing - review & editing.

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findings, and conclusions or recommendations expressed in this material are those of the author(s) and do not necessarily reflect the views of the National Science Foundation. Scientific collection in Panama is done with permission from landowners and prior approval of MiAmbiente, Panama's environmental authority (permit number: SE/A-17-18), and the Institutional Animal Care and Use Committee of the Smithsonian Tropical Research Institute (IACUC permit: 2018-0116-2021) and the University of Tennessee (IACUC permit: 2573). We thank the Rosvall lab for feedback, Elizabeth George for assisting with testosterone assay validation, Evan Buck for assistance in the field, Matt Fuxjager and Meredith Miles for assistance importing and storing samples, the STRI Bird Collection for preparing taxidermic mounts, Mae Berlow for the jacana illustrations, and Elizabeth Derryberry for conceptual design and funding acquisition.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ygcen.2020.113444>.

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