

# Risk Factors for Recurrent *Staphylococcus aureus* Bacteremia

Seong-Ho Choi,<sup>1,2</sup> Michael Dagher,<sup>1</sup> Felicia Ruffin,<sup>1</sup> Lawrence P. Park,<sup>1,3</sup> Batu K. Sharma-Kuinkel,<sup>1</sup> Maria Souli,<sup>1,4</sup> Alison M. Morse,<sup>5,6</sup> Emily M. Eichenberger,<sup>1</sup> Lauren Hale,<sup>1</sup> Celia Kohler,<sup>1</sup> Bobby Warren,<sup>1</sup> Brenda Hansen,<sup>1,7</sup> Felix Mba Medie,<sup>1</sup> Lauren M. McIntyre,<sup>5,6</sup> and Vance G. Fowler Jr.<sup>1,4</sup>

<sup>1</sup>Department of Medicine, Duke University Medical Center, Durham, North Carolina, USA, <sup>2</sup>Division of Infectious Diseases, Department of Internal Medicine, Chung-Ang University Hospital, Chung-Ang University College of Medicine, Seoul, South Korea, <sup>3</sup>Duke Global Health Institute, Duke University, Durham, North Carolina, USA, <sup>4</sup>Duke Clinical Research Institute, Durham, North Carolina, USA, <sup>5</sup>Departments of Molecular Genetics and Microbiology, University of Florida, Gainesville, Florida, USA, <sup>6</sup>University of Florida Genetics Institute University of Florida, Gainesville Florida, USA, and <sup>7</sup>Pediatric Gastroenterology, University of North Carolina, Chapel Hill, North Carolina, USA

**Background.** To understand the clinical, bacterial, and host characteristics associated with recurrent *Staphylococcus aureus* bacteremia (R-SAB), patients with R-SAB were compared to contemporaneous patients with a single episode of SAB (S-SAB).

**Methods.** All SAB isolates underwent *spa* genotyping. All isolates from R-SAB patients underwent pulsed-field gel electrophoresis (PFGE). PFGE-indistinguishable pairs from 40 patients underwent whole genome sequencing (WGS). Acute phase plasma from R-SAB and S-SAB patients was matched 1:1 for age, race, sex, and bacterial genotype, and underwent cytokine quantification using 25-analyte multiplex bead array.

**Results.** R-SAB occurred in 69 (9.1%) of the 756 study patients. Of the 69 patients, 30 experienced relapse (43.5%) and 39 reinfection (56.5%). Age, race, hemodialysis dependence, presence of foreign body, methicillin-resistant *Staphylococcus aureus*, and persistent bacteremia were individually associated with likelihood of recurrence. Multivariate risk modeling revealed that black hemodialysis patients were nearly 2 times more likely (odds ratio [OR] = 9.652 [95% confidence interval [CI], 5.402–17.418]) than white hemodialysis patients (OR = 4.53 [95% CI, 1.696–10.879]) to experience R-SAB. WGS confirmed PFGE interpretations in all cases. Median RANTES (regulated on activation, normal T cell expressed and secreted) levels in acute phase plasma from the initial episode of SAB were higher in R-SAB than in matched S-SAB controls ( $P = .0053$ , false discovery rate < 0.10).

**Conclusion.** This study identified several risk factors for R-SAB. The largest risk for R-SAB is among black hemodialysis patients. Higher RANTES levels in R-SAB compared to matched controls warrants further study.

**Keywords.** *Staphylococcus aureus*; bacteremia; recurrence; whole genome sequencing; health disparity.

*Staphylococcus aureus* bacteremia (SAB) is a common and potentially lethal infection [1]. Approximately 2–20% of patients with an initial episode of SAB will develop a recurrent *S. aureus* bacteremia (R-SAB) after the resolution of the initial infection [2–4]. Although several previous studies have sought to identify risk factors for R-SAB [5–11], none have simultaneously considered clinical, bacterial, and host inflammatory characteristics.

In the current investigation, we used a large prospective cohort of patients with SAB to identify clinical characteristics associated with R-SAB as compared with patients who had only a single episode of SAB (S-SAB). Next, we genotyped the paired bacterial isolates from the repeat SAB episodes to differentiate

patients with recurrent SAB due to a persistent source (relapse) from patients with a new episode of SAB (reinfection) using 3 molecular techniques. Finally, we tested the possibility that patients with R-SAB exhibited fundamental differences in their response to *S. aureus* that predisposed them to recurrence by comparing the cytokines from acute phase plasma of matched patients with R-SAB and S-SAB.

## METHODS

### Study Population

Since September 1994, the SAB Group Prospective Cohort Study (SABG-PCS) has prospectively enrolled all eligible adult (age  $\geq 18$  years) hospitalized nonneutropenic (absolute neutrophil count  $> 1 \times 10^9/L$ ) patients with monomicrobial SAB at Duke University Medical Center. Demographics, past medical history, history of surgery within the previous 30 days, site of acquisition of SAB, APACHE (Acute Physiology and Chronic Health Evaluation) II score calculated on the day of the index positive blood culture, metastatic complications, and patient outcome were collected on a standardized case report form and entered into an electronic database. Using the SABG-PCS data

Received 1 September 2019; editorial decision 20 May 2020; accepted 12 June 2020; published online June 21, 2020.

Correspondence: V. G. Fowler, Jr., Rm 185 Hanes Bldg, 315 Trent Dr, Division of Infectious Diseases, Department of Medicine, Box 102359 Duke University Medical Center, Durham, NC 27710 (vance.fowler@duke.edu).

Clinical Infectious Diseases® 2021;72(11):1891–9

© The Author(s) 2020. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com.  
DOI: 10.1093/cid/ciaa801

between January 2008 and May 2015, patients with SAB were selected. Patients included in this study are part of a larger cohort discussed elsewhere [12] and were previously presented in part at IDWeek 2018 [13]. This study was approved by the Duke Institutional Review Board. Patients (or legal representative) provided written informed consent.

### Definitions

R-SAB was defined as a second episode of SAB after the resolution of the first episode occurring at least 14 days from the date of the last positive blood culture of the first episode [14]. SAB was categorized as community-acquired, healthcare-associated, or hospital-acquired, as described elsewhere [15]. A foreign body was defined as any device that was inserted for an extended period of time (eg, tunneled intravascular catheter, synthetic intravascular graft, arthroplasty, orthopedic hardware, prosthetic valves, and cardiac devices). Persistent bacteremia was defined as  $\geq 5$  days of positive blood cultures after appropriate treatment was initiated. Patients were considered to have hematogenous metastatic infections from their bacteremia if they developed any of the following conditions during their hospitalization for SAB after the first positive culture: infective endocarditis, vertebral osteomyelitis, septic arthritis, septic emboli, septic thrombophlebitis, metastatic abscess, or other deep tissue abscess (ie, epidural or psoas abscess).

### Laboratory Studies

Bacterial isolates were speciated by the Duke Clinical Microbiology Laboratory using standard techniques. Minimum inhibitory concentration values were determined using an automated broth microdilution method (MicroScan WalkAway plus System, Beckman Coulter, Brea, CA, USA). Methicillin susceptibility was defined according to Clinical and Laboratory Standards Institute (CLSI) guidelines.

### Pulsed-field Gel Electrophoresis (PFGE)

PFGE was performed for all *S. aureus* blood isolates from patients with R-SAB, as previously described [16]. PFGE patterns were compared between isolates for each patient with R-SAB by visual inspection and interpreted as either “indistinguishable” or “discordant” using established guidelines [17] by 2 experienced investigators (B. K. S.; B. W.) blinded to clinical category of the source patient.

### Defining and Differentiating Relapse Versus Reinfection in Patients With R-SAB

R-SAB patients with PFGE-discordant isolate pairs were defined as having reinfection. In R-SAB patients with PFGE-indistinguishable isolate pairs, the time between the first episode of SAB and the subsequent recurrent episode ( $\Delta T$ ) was used to differentiate relapse from reinfection [18, 19]. Patients in whom the PFGE-indistinguishable pair of isolates occurred at  $\Delta T \geq 150$  days apart were defined as having reinfection with

an identical strain. By contrast, patients in whom the PFGE-indistinguishable pair of isolates occurred  $< 150$  days apart were defined as having *relapse*.

### Spa Genotyping

The initial *S. aureus* bloodstream isolate from all study patients underwent *spa* typing as described elsewhere [20]. Assignment of *spa* types was performed using Ridom StaphType™ (Ridom GmbH, Wurzburg, Germany). *Spa* types were clustered into *spa* clonal complexes (*spa*-CCs) using the Based Upon Repeat Pattern (BURP) algorithm at a cost setting of  $\leq 4$  and excluding *spa* types with  $< 5$  repeats.

### Whole Genome Sequencing

Whole genome Illumina sequencing (paired end, read length 150 bp) was carried out on 40 isolate pairs (80 isolates total). One sample failed sequencing; it and its pair were not considered further (see [Supplementary methods](#)).

### Cytokine Analysis

Acute phase plasma was obtained from consenting study participants within  $\sim 3$  days after initial positive blood cultures for SAB. To conduct cytokine analysis, 21 patients with R-SAB were matched 1:1 to patients with S-SAB based upon age, sex, race, and *spa*-CC of the bloodstream *S. aureus* isolate. For 18 of the 21 samples, all 4 variables were matched. The remaining 3 patients were matched on as many of these variables as possible. Plasma cytokine and chemokine concentrations were assayed using a 25-analyte multiplex bead array (GM-CSF, Eotaxin, interferon [IFN]- $\alpha$ , induced protein [IP]-10, IFN- $\gamma$ , monocyte chemoattractant protein [MCP]-1, interleukin [IL]-1 $\beta$ , monokine induced by interferon- $\gamma$  [MIG], IL-1RA, macrophage inflammatory protein [MIP]-1 $\alpha$ , IL-2, MIP-1 $\beta$ , IL-2R, RANTES [regulated on activation, normal T cell expressed and secreted], IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-12 [p40/p70], IL-13, IL-15, IL-17, tumor necrosis factor [TNF]- $\alpha$ ; Invitrogen; Carlsbad) prepared according to the manufacturer's recommended protocol and read using a Bio-Plex 200 suspension array reader (Bio-Rad). Data were analyzed using Bio-Plex manager software v6.1 (Bio-Rad; Hercules, CA, USA).

### Statistical Analysis

Continuous variables were compared using the Wilcoxon 2-sample rank-sum test and categorical variables using Fisher exact test. The cytokine levels of baseline plasma were compared using Wilcoxon signed rank test between matched pairs. Logistic regression analysis was performed to identify independent risk factors for R-SAB. Variables with  $P < .10$  on univariate analysis were included in a multivariate logistic regression analysis. Where variables were strongly correlated, the correlation structure was accounted for. Multiple comparisons adjustment was made as appropriate with false discovery rate (FDR) 10%.

## RESULTS

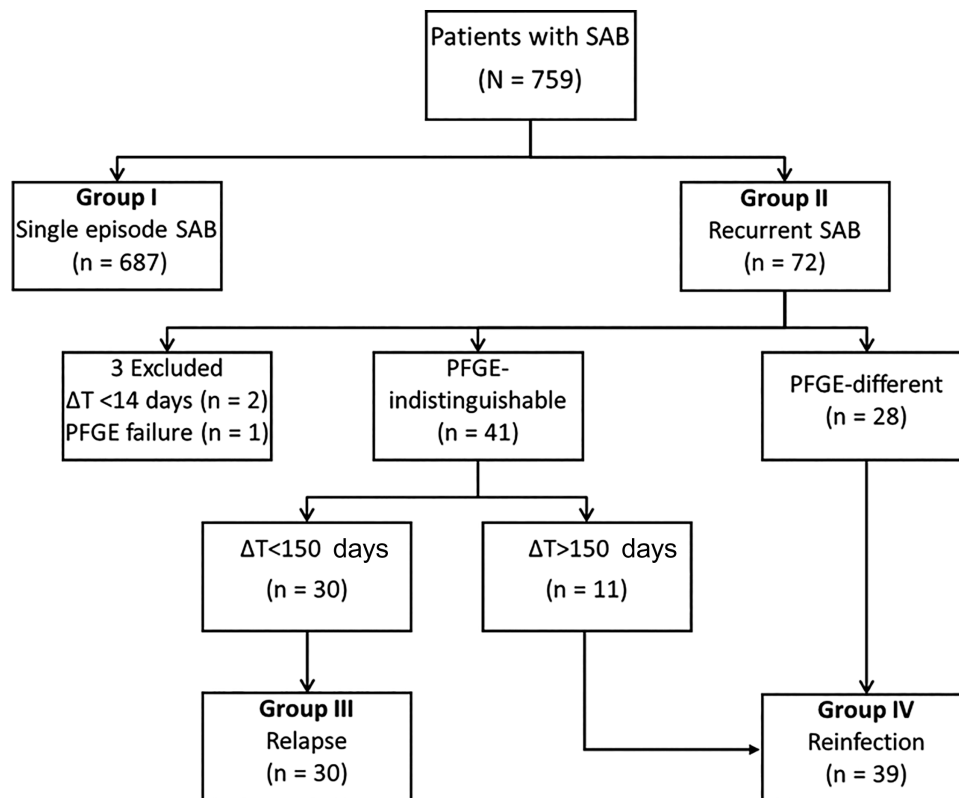
### Inclusion of the Study Patients and Differentiation Between Relapse and Reinfection

During the 7-year study period, a total of 759 patients with SAB were enrolled. Of these, 687 patients experienced S-SAB and 72 patients (9.5%) had R-SAB. After the exclusion of 2 patients whose  $\Delta T$  was < 14 days and 1 patient whose isolates failed to show any PFGE band in repeated tests, 69 patients with R-SAB were included in the study analyses. Of these 69 patients, 55 experienced a single recurrence (2 total episodes of SAB) and 14 experienced multiple recurrences ( $\geq 3$  episodes of SAB). Patients with 1 recurrence did not differ significantly from those with multiple recurrences (Table S1). For patients with multiple recurrences, only the first and second episodes were analyzed. Overall, the median duration of  $\Delta T$  until the first recurrence was 143 days (interquartile range [IQR] 77d – 354d). PFGE performed on sequential isolates corresponding to the first recurrence resulted in 41 indistinguishable pairs and 28 different pairs (Figure S4). All sequential pairs with  $\Delta T < 60$  days were caused by indistinguishable strains (eg, relapse). The first occurrence of R-SAB due to a PFGE-different strain appeared during the third month, and 75% of the sequential pairs with different PFGE profiles (eg, reinfection) occurred at  $\Delta T \geq 150$  days. For this reason, if a sequential pair with indistinguishable

PFGE profiles occurred at  $\Delta T \geq 150$  days, it was interpreted as reinfection with an identical strain for the purposes of the present analysis. By contrast, PFGE-indistinguishable pairs obtained < 150 days apart were classified as relapse. PFGE-discordant pairs were all considered to be reinfections. Using this definition, the study participants were classified into 2 groups: relapse (30 patients; 43.5%) and reinfection (39 patients; 56.5%) (Figure 1).

### Patients With R-SAB Versus Patients With S-SAB

As compared to patients with S-SAB, R-SAB patients were significantly younger (56y vs 61y,  $P = .0003$ ); more frequently black (63.8% vs 29.8%,  $P < .0001$ ) and hemodialysis dependent (55.1% vs 15.6%,  $P < .0001$ ); more likely to have a foreign body (82.6% vs 59.7%,  $P = .0001$ ), more likely to exhibit persistent bacteremia (39.1% vs 24.7%,  $P = .0138$ ), and be infected with methicillin-resistant *S. aureus* (MRSA) (56.5% vs 43.7%,  $P = .0429$ ) (Table 1). Conversely, patients with S-SAB were more likely to have a diagnosed neoplasm (25.5% vs 11.6%,  $P = .008$ ) and to have had surgery within the 30 days preceding infection (23.7% vs 11.6%,  $P = .0229$ ). Surgery within 30 days, presence of a foreign body, age, and APACHE II scores showed significant pairwise correlation. In addition, there was a strong relationship between race and hemodialysis dependence. To explore this



**Figure 1.** Flow of classification of study patients based on PFGE and cutoff value of the time between the first episode of SAB and the subsequent episode of recurrent SAB ( $\Delta T$ ). Abbreviations: PFGE, pulsed-field gel electrophoresis; SAB, *S. aureus* bacteremia.

**Table 1. Clinical Characteristics of Patients With Single Episode of *Staphylococcus aureus* Bacteremia (SAB) (Group I), Patients With Recurrent SAB (Group II), Patients With Relapse of SAB (Group III), and Patients With Reinfection of SAB (Group IV)**

Characteristics	Group I Single episode (N = 687)	Group II Recurrence (N = 69)	P value, I vs II	Group III Relapse (N = 30)	P value I vs III	Group IV Reinfection (N = 39)	P value I vs IV	P value III vs IV
<b>Demographics</b>								
Median age (IQR)	61.0 (50.0–71.0)	56.0 (40.0–63.0)	<b>.0003</b>	58.0 (51.0–63.0)	.0615	46.0 (35.0–64.0)	<b>.001</b>	.3061
Male sex	419 (61.0)	40 (58.0)	.70	23 (76.7)	.0882	17 (43.6)	<b>.0423</b>	<b>.0073</b>
Race <sup>a</sup>			<b>&lt;.0001</b>		<b>.0080</b>		<b>&lt;.0001</b>	.2021
White	460(67.2)	24 (34.8)		13 (43.3)		11 (28.2)		
Black	205 (29.8)	44 (63.8)		16 (53.3)		28 (71.8)		
Other	20 (2.9)	1 (1.4)		1 (3.3)		0		
Acquisition <sup>b</sup>			.43		.1001		.6466	.1387
Hospital-acquired	120 (17.5)	8 (11.8)		1 (3.3)		7 (17.9)		
Healthcare-associated	473 (68.9)	52 (76.5)		23 (76.7)		29 (74.4)		
Community-acquired	94 (13.7)	8 (11.8)		5 (16.7)		3 (7.7)		
<b>Underlying disease/condition</b>								
Diabetes mellitus	279 (40.6)	34 (49.3)	.1995	16 (53.3)	.19	18 (46.2)	.5070	.6307
Hemodialysis dependence	107 (15.6)	38 (55.1)	<b>&lt;.0001</b>	16 (53.3)	<b>&lt;.0001</b>	22 (56.4)	<b>&lt;.0001</b>	.8123
Injection drug use	28 (4.1)	1 (1.4)	.51	0	.62	1 (2.56)	1.00	1.00
Neoplasm	175 (25.5)	8 (11.6)	<b>.008</b>	6 (20.0)	.6681	2 (5.13)	<b>.0019</b>	.0702
Transplantation	59 (8.6)	6 (8.7)	1.00	4 (13.3)	.3253	2 (5.13)	.7641	.3920
Steroid use	163 (23.7)	10 (14.5)	.098	6 (20.0)	.8264	4 (10.3)	.0517	.3121
HIV infection <sup>c</sup>	16 (2.3)	3 (4.4)	.24	2 (6.7)	.1636	1 (2.56)	.6146	.5712
Foreign body <sup>d</sup>	410 (59.7)	57 (82.6)	<b>.0001</b>	26 (86.7)	<b>.0034</b>	31 (79.5)	<b>.0172</b>	.5316
Surgery within 30 days <sup>e</sup>	163 (23.7)	8 (11.6)	<b>.0229</b>	3 (10.0)	.0814	5 (12.8)	.1226	1.00
<b>Clinical features of SAB</b>								
Median APACHE II score (IQR)	16.0 (12.0 – 22.0)	15.0 (12.0 – 18.0)	.07	15.0 (12.0 – 19.0)	.18	16.05 (12.0 – 18.0)	.30	.7520
Persistent bacteremia	170 (24.7)	27 (39.1)	<b>.0138</b>	14 (46.7)	<b>.0103</b>	13 (33.3)	.2551	.3229
Infective endocarditis <sup>e</sup>	122 (17.8)	15 (21.7)	.3590	5 (16.7)	1.00	10 (25.6)	.2011	.3829
Metastatic abscess <sup>e</sup>	78 (11.4)	12 (17.4)	.1013	7 (23.3)	<b>.0384</b>	5 (12.82)	.5853	.3426
Metastatic arthritis <sup>e</sup>	54 (7.9)	4 (5.8)	.81	3 (10.0)	.48	1 (2.56)	.51	.3148
Septic thrombophlebitis <sup>e</sup>	40 (5.8)	5 (7.2)	.4658	3 (10.0)	.2644	2 (5.13)	1.00	.6480
<b>Microbiologic characteristics</b>								
Methicillin resistance	300 (43.7)	39 (56.5)	<b>.0429</b>	22 (73.3)	<b>.0022</b>	17 (43.6)	1.00	<b>.0160</b>

Abbreviations: HIV, human immunodeficiency virus; IQR, interquartile range.

<sup>a</sup>Data missing for 2 patients from Group I; P-values reported are for black vs white.

<sup>b</sup>Data missing for 1 patient from Group II, originating from Group III.

<sup>c</sup>Data missing for 1 patient from Group III.

<sup>d</sup>Data missing for 1 patient from Group I.

<sup>e</sup>Data missing for 6 patients from Group II, 2 originating from Group III, and 4 from group IV.

further, we constructed a multilevel variable to capture potential interactions between race and hemodialysis dependence by defining 3 levels: “black and hemodialysis dependent,” “white and hemodialysis dependent,” and “not hemodialysis dependent.”

A multivariate logistic regression model was generated based on predictors that had  $P < .10$  in initial models (Table S2) and using the race and hemodialysis combination variable. Variables were eliminated based on a lack of independent predictive value with clinical relevance used to judge selection among correlated variables. In the final model, the combination variable of race/hemodialysis dependence was an independent risk factor for R-SAB ( $P < .001$ ). Blacks on hemodialysis were at the highest risk with an adjusted odds ratio of 9.652 (95% confidence interval [CI], 5.402–17.418), followed by hemodialysis

dependent whites (4.53 [95% CI, 1.696–10.879]) compared to nonhemodialysis dependent controls. An APACHE II score above median value was also a significant predictor of recurrence (1.869 [95% CI, 1.081–3.271],  $P = .0263$ ) (Table 2).

Survival, mortality, and recurrence are part of the criteria that define S-SAB versus R-SAB study design and could therefore only be evaluated in the S-SAB group. Within S-SAB, blacks and whites had virtually identical survival (68.8% and 67.6%, respectively) and attributable mortality (18.1% and 19.4%) rates.

#### Risk Factors for Relapse and Reinfection R-SAB

As compared to patients with S-SAB, blacks (compared to whites), hemodialysis dependence, and the presence of an



**Table 2. Multiple Logistic Regression Model Predicting Recurrent *Staphylococcus aureus* Bacteremia**

Risk Factor	Final Regression Model C = 0.736	
	Adjusted Odds Ratio (95% CI)	P-value
Dialysis and race		< .0001
Dialysis dependent blacks	9.652 (5.402–17.418)	
Dialysis dependent whites	4.53 (1.696–10.879)	
APACHE score above median	1.869 (1.081–3.271)	.0263

Abbreviation: CI, confidence interval.

indwelling foreign body were significantly associated with both relapse and reinfection (Table 1). Persistent bacteremia (46.7% vs 24.7%,  $P = .0103$ ), methicillin resistance (73.3% vs 43.7%,  $P = .0022$ ), and metastatic abscess (23.3% vs 11.4%,  $P = .0384$ ) were more frequent in patients with relapsed SAB than patients with S-SAB. Patients with reinfection were younger (46y vs 61y;  $P = .0010$ ), more likely to be female (56.4% vs 39.0%,  $P < .0423$ ) and less likely to have a neoplasm (5.13% vs 25.5%,  $P = .0019$ ) compared to patients with S-SAB.

#### Isolate Genotypes

We sought to understand whether whole genome sequencing (WGS) provided additional insights not apparent in the PFGE analysis. The paired isolates selected for sequencing all had highly related or identical PFGE profiles. WGS data revealed that these paired isolates shared MLST ST types and differed by  $\leq 100$  single-nucleotide polymorphisms (SNPs). This finding suggests that these highly related isolates either arise from a single persistent patient source of infection or from multiple unrelated infections with a single successful clone [21]. To see if we could separate these possibilities, we compared all possible pairs of isolates between different individuals. A small subset had fewer than 200 SNPs ( $n = 72$  out of 1040 pairs of isolates). These 72 pairs are combinations of isolates originating from a few patients. We compared the PFGE among these patients and found that the patterns are a visual match, indicating that the SNP counts agree with PFGE (Figure S5). This implies that we have identified a few successful clones in our sample. These successful clones are a small subset of the most frequently occurring Spa-types in this population: MLST ST105/5 (Spa-CC002) and ST8 (Spa-CC008). We compared the distribution of SNPs in the PFGE matched isolates from different patients to the PFGE matched isolate pairs from the same individual (Figure 2). The distributions are different with fewer SNP differences in the pairs from the same patient (median = 31 vs 178,  $P < .0001$ ). However, the range observed is similar in both cases. This similarity indicates that the number of SNPs alone is insufficient to distinguish between independent

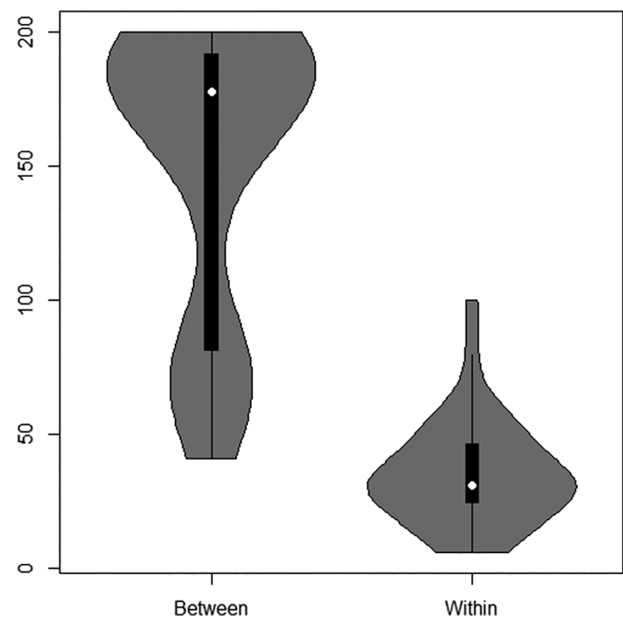
reinfection from a successful clone, reinfection from a clone in an external reservoir, or a relapse of an incompletely resolved infection within a single patient. We conclude that the amount of SNP variation by WGS is consistent with the PFGE and that no novel insights were provided by the WGS in these data.

#### Genotype of *S. aureus* Isolates in Patients With R-SAB and S-SAB

The most common *spa*-CCs in our sample were CC2 and CC8 (Table 3). Although CC8 was more frequently observed in the relapse group (40%), this was not significantly different from the S-SAB group (29.7%,  $P = .1341$ ) or the reinfection group (25.6%,  $P = .8411$ ). The distribution of CC2 was similar in all groups.

#### Acute Phase Plasma Cytokines in Patients With R-SAB and S-SAB

Next, we evaluated acute phase plasma cytokine levels from the initial episode of bacteremia in patients with R-SAB and S-SAB. Twenty-one patients with R-SAB were selected based on the availability of their baseline (first episode) plasma. Among the 25 cytokines tested (Figure S6), only RANTES exhibited significant differences between patients with R-SAB and S-SAB (Figure 3). We define  $\Delta$ RANTES to be the difference in RANTES values between a case of R-SAB and its matched S-SAB control ( $\Delta$ RANTES = RANTES R-SAB – RANTES control).  $\Delta$ RANTES was positive for 18 of the 21 cases of R-SAB (Figure 3A; WRS  $P = .0053$ , false discovery rate  $< 0.10$ ) [22]. Three cases of R-SAB with a negative  $\Delta$ RANTES all came



**Figure 2.** Distribution of the number of SNPs for isolate pairs between individuals with fewer than 200 SNPs (between) compared to pairs of isolates from the same individual with a PFGE match (within). Supplementary Figure 5 indicates that the isolate pairs between individuals are a visual PFGE match. Abbreviations: PFGE, pulsed-field gel electrophoresis; SNP, single-nucleotide polymorphism.

**Table 3. Distribution of *Spa*-clonal Complexes (CC) of Patients for Each Categorization of SAB**

<i>Spa</i> -clonal Complex	Group I Single Episode (n = 687)	Group II Recurrence (n = 69) <sup>a</sup>	<i>P</i> -value, I vs II	Group III Relapse (n = 30) <sup>b</sup>	<i>P</i> -value, I vs III	Group IV Reinfection (n = 39) <sup>c</sup>	<i>P</i> -value, I vs IV
CC2	208 (30.3)	21 (30.4)	.2922	8 (26.7)	.8276	13 (33.3)	.1695
CC8	204 (29.7)	22 (31.9)	.1341	12 (40.0)	.0736	10 (25.6)	.8411
Other <sup>d</sup>	275 (40.0)	13 (18.8)		5 (16.7)		8 (20.5)	

Abbreviation: SAB, *Staphylococcus aureus* bacteremia.

<sup>a</sup>Data missing from 13 patients.

<sup>b</sup>Data missing for 5 patients.

<sup>c</sup>Data missing for 8 patients.

<sup>d</sup>This category includes *spa*-types CC12, CC148, CC164, CC189, CC216, CC4, CC78/81, CC84, nontypeable, and not available; statistical comparison of these subgroups was not performed due to low cell counts.

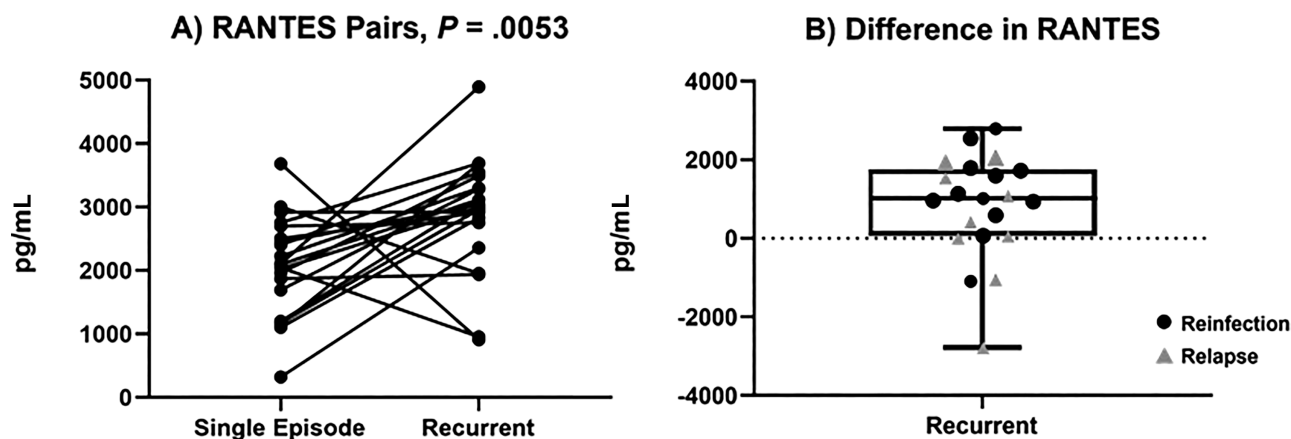
from perfectly matched pairs. RANTES values were not significantly correlated (range  $-0.13$  to  $0.26$ ; median  $-0.02$ ) with any other cytokines, and although  $\Delta$ RANTES was positive for most matched pairs, equivalent metrics for the majority of other cytokines were negative.

## DISCUSSION

The prevalence of R-SAB in our study was  $\sim 9\%$ , similar to earlier reports [6, 7]. However, the documented prevalence of R-SAB varies widely (Table S3). This variation is influenced by study design, definitions of recurrence, and the proportion of MRSA. For example, recurrence rates varied from 7.1% in a study where MRSA accounted for  $< 1\%$  of the sample [11] to  $> 20\%$  in a study limited to MRSA infections [9]. Kreisel et al reported a high rate of R-SAB ( $\sim 17\%$ ), but excluded patients who died before the completion of antibiotics [8]. Conversely, Albertson et al reported a lower R-SAB rate of 6.3%, but excluded recurrent cases more than 180 days after the initial episode of SAB [5]. Thus, our rate of R-SAB is consistent with what has been previously reported in the literature.

Blacks comprised  $\sim$  one-third of the entire study cohort. In the multivariate model, black hemodialysis patients were  $\sim 2$  times more likely than white hemodialysis counterparts to develop R-SAB. This is consistent with previous research showing that there is a substantial health disparity in the incidence of invasive MRSA infections compared to white patients [23, 24].

Socioeconomic status (SES), factors such as poverty, crowding, and the availability and affordability of medical care have been shown to contribute [25–27] to health disparities in SAB. Indeed, See et al reported that after controlling for socioeconomic factors in a mediation analysis, there was no disparity between black and white patients in MRSA infection [25]. This is supported by data from outside the United States, suggesting that lower SES is associated with higher rates of MRSA infection [26, 27]. In addition, there is some evidence to suggest a relationship between SES and host colonization with *S. aureus* [28, 29]. For example, Freitas et al found self-reported black race as a risk factor for colonization with a highly virulent strain of MRSA (OR, 1.81 [95% CI, 1.38–2.38]) [30]. Differences in allele frequencies within HLA Class II and other genetic factors



**Figure 3.** A, Comparison of baseline plasma RANTES level between patients with recurrent SAB) and age/sex/race/genotype-matched patients with a resolving single episode of SAB. B, Difference in RANTES values between recurrent SAB and their matched single SAB. We also examined a more conservative set of fewer than 100 SNPs. Here the median for the “across individual” set was 74 and still significantly different from the “same individual” pairs ( $P < .0001$ ). Abbreviations: RANTES, regulated on activation, normal T cell expressed and secreted; SAB, *Staphylococcus aureus* bacteremia; SNP, single-nucleotide polymorphism.

controlling innate immunity also influence host susceptibility to *S. aureus* infection in both black and white subjects [31, 32].

In our study, black race in combination with hemodialysis dependence conferred significant risk of R-SAB, with an OR of over 9. These patients were twice as likely to have R-SAB as white hemodialysis dependent patients (OR, 4.5). This is consistent with previous studies. In a study that evaluated the impact of race on risk for invasive MRSA, blacks were at an increased risk for both healthcare-associated MRSA infections in general (adjusted rate ratio [aRR], 3.84 [95% CI, 2.94–5.01]) and hemodialysis-associated infection in particular (aRR, 1.83 [95% CI, 1.72–1.96]) [23]. In the United States, non-Hispanic blacks have the highest lifetime risk of developing end-stage renal disease (ESRD) [33] and are subsequently at an increased risk of becoming hemodialysis-dependent [34]. In addition, Saunders et al demonstrated that hemodialysis-dependent blacks are less likely to use hemodialysis facilities that are rated highly by federal quality reporting programs [35].

Although hemodialysis dependent blacks were more likely to experience R-SAB than white hemodialysis dependent patients, the 2 groups had similar APACHE II scores, rates of metastatic infections, and persistent bacteremia. In addition, survival and SAB-attributable mortality for black and white patients in the S-SAB group were virtually identical. These findings are consistent with a previous study suggesting that race did not influence risk-adjusted mortality in patients with sepsis despite a health disparity in the incidence [36].

Higher APACHE II scores were also a significant predictor of R-SAB in our multivariate model. This association between measures of illness severity and risk of recurrence has been previously demonstrated [5]. APACHE II scores are widely used, easily interpreted, and include estimates for both acute and chronic illness severity estimates, which can reduce the number of variables included in a multivariable model [8, 11]. Thus, we used APACHE II in our final multivariable model as a parsimonious strategy to address patient comorbidity. Using this strategy, we found that patients with higher levels of acute illness severity and more comorbid conditions, represented by APACHE II > 15, were 1.869 (95% CI, 1.081–3.271) times more likely to develop R-SAB.

R-SAB includes bacteremia due to both relapse and reinfection. Differentiating these syndromes is clinically important, but often challenging. Relapsing SAB typically suggests an inability to eradicate an established *S. aureus* infection or potentially an additional episode of bacteremia with the same bacterial strain. Nasal carriage of *S. aureus* is a significant risk factor for developing SAB [37, 38] and is often the source of the isolate causing bacteremia [39]. In our study and previous research, persistent bacteremia [5], metastatic abscesses, and MRSA [8] were associated with relapse. This is due in part to their association with infections involving high bacterial load, unremoved foci, and/or insufficient antimicrobial treatment [3, 6]. On the other

hand, new episodes of reinfection in the same host suggest an increased susceptibility to recurrent bouts of *S. aureus* infection. This increased susceptibility may be due to a medical (eg, long-term catheter), environmental (injection drug use), or genetic condition. In our study, reinfection was associated with younger age, a finding that aligns with previous reports [5].

Despite their increased risk to develop R-SAB, hemodialysis-dependent black patients were no more likely than non-hemodialysis patients to relapse compared to reinfection ( $P = .20$ ). This finding suggests that the observed health disparity reflects a general susceptibility to recurrence as opposed to an inability to successfully clear infection or exposure to an untreated reservoir. This interpretation is supported by the similar time to recurrence observed in black hemodialysis-dependent patients, white hemodialysis-dependent patients, and nonhemodialysis patients ( $P = .60$ ).

Host inflammatory response was associated with R-SAB. The median level of chemokine ligand 5 (CCL5), also known as RANTES, was significantly higher in the baseline plasma of the 21 patients with R-SAB than in the baseline plasma of 21 matched subjects with S-SAB. There was no association between race and RANTES levels. RANTES is a chemotactic cytokine that recruits leukocytes to infection sites [40]. In a recent study investigating cytokine responses to SAB, levels of RANTES rose significantly in SAB patients by day 7 and remained persistently elevated up to 14 days in patients with complicated SAB [41]. Although the biological mechanism is yet to be determined, our findings suggest that RANTES may play a role in the host immune response to SAB. Further study is needed to validate this discovery and understand the underlying mechanism.

This study has limitations. First, we categorized PFGE-indistinguishable strains into relapse and reinfection based on the time point at which the second episode of SAB occurred. Thus, we may have inaccurately classified patients who relapsed after 149 days as having experienced a reinfection. Second, patients may have sought medical attention at other hospitals for their recurrent episode of SAB. Third, the impact of antimicrobial and surgical therapy on the occurrence of R-SAB was not evaluated in this study. Fourth, it is not possible to compare outcomes among patients with R-SAB and S-SAB due to the definition of these 2 groups. Finally, this is a single-center study with a small sample size for certain events and measurements.

In conclusion, the effects of SAB can be compounded by recurrent episodes in some patients. Our study demonstrated a racial health disparity in risk for R-SAB that is consistent with previous literature on racial disparities in SAB. Further research is needed to fully elucidate the mechanisms driving this disparity. In addition, illness severity increases the likelihood of recurrence. Finally, in a small matched pair cohort, RANTES at baseline was higher in patients who went on to develop R-SAB than in patients with S-SAB, and this trend was not associated with race. Further study on the mechanisms of R-SAB are

warranted. In particular, adequately powered studies of black patients are critical.

### Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

### Notes

**Funding.** This research was supported in part by the following grants from the National Institutes of Health: U01-AI124319, R01-AI068804, R01-GM128193, U2-CE030167. Dr. Fowler was supported by National Institutes of Health Grant K24-AI093969. Biomarker profiling was performed under the management of Dr Andrew N. Macintyre and direction of Dr Gregory D. Sempowski in the Immunology Unit of the Duke Regional Biocontainment Laboratory (RBL), which received partial support for construction from the National Institutes of Health, National Institute of Allergy and Infectious Diseases (UC6-AI058607).

**Potential conflicts of interest.** V. G. F. reports grant/ research support from the National Institutes of Health, MedImmune, Cerexa/Forest/Actavis/Allergan, Pfizer, Advanced Liquid Logics, Theravance, Novartis, Cubist/Merck; Medical Biosurfaces; Locus; Affinergy; Contrafect; Karius; Gtenentech, Regeneron, BasileaPaid Consultant: Pfizer, Novartis, Galderma, Novadigm, Durata, Debiopharm, Genentech, Amphlphi Biosciences, Integrated Biotherapeutics, C3J, Achaogen, Affinium, Medicines Co., Cerexa, Tetrphase, Trius, MedImmune, Bayer, Theravance, Cubist, Basilea, Affinergy, Janssen, xBiotech, Contrafect, Regeneron, Basilea, Destiny. Membership: Merck Co-Chair V710 Vaccine. Educational fees: Green Cross, Cubist, Cerexa, Durata, Theravance; Debiopharm. Royalties from UpToDate. Patent pending for host gene expression signature diagnostic for sepsis. All other authors have no potential conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

### References

1. Fowler VGJ, Holland TL. Clinical approach to *Staphylococcus aureus* bacteremia in adults. In: Lowy FD, ed. UpToDate. Waltham, MA: UpToDate Inc.
2. Fowler VG Jr, Olsen MK, Corey GR, et al. Clinical identifiers of complicated *Staphylococcus aureus* bacteremia. *Arch Intern Med* 2003; 163:2066–72.
3. Hawkins C, Huang J, Jin N, Noskin GA, Zembower TR, Bolon M. Persistent *Staphylococcus aureus* bacteremia: an analysis of risk factors and outcomes. *Arch Intern Med* 2007; 167:1861–7.
4. Tong SY, Davis JS, Eichenberger E, Holland TL, Fowler VG Jr. *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clin Microbiol Rev* 2015; 28:603–61.
5. Albertson J, McDanel JS, Carnahan R, et al. Determination of risk factors for recurrent methicillin-resistant *Staphylococcus aureus* bacteremia in a Veterans Affairs healthcare system population. *Infect Control Hosp Epidemiol* 2015; 36:543–9.
6. Chang FY, Peacock JE Jr, Musher DM, et al. *Staphylococcus aureus* bacteremia: recurrence and the impact of antibiotic treatment in a prospective multicenter study. *Medicine (Baltimore)* 2003; 82:333–9.
7. Fowler VG Jr, Kong LK, Corey GR, et al. Recurrent *Staphylococcus aureus* bacteremia: pulsed-field gel electrophoresis findings in 29 patients. *J Infect Dis* 1999; 179:1157–61.
8. Kreisel K, Boyd K, Langenberg P, Roghmann MC. Risk factors for recurrence in patients with *Staphylococcus aureus* infections complicated by bacteremia. *Diagn Microbiol Infect Dis* 2006; 55:179–84.
9. Liao CH, Lai CC, Chen SY, Huang YT, Hsueh PR. Strain relatedness of methicillin-resistant *Staphylococcus aureus* isolates recovered from patients with repeated bacteraemia. *Clin Microbiol Infect* 2010; 16:463–9.
10. Walker TM, Bowler IC, Bejon P. Risk factors for recurrence after *Staphylococcus aureus* bacteraemia: a retrospective matched case-control study. *J Infect* 2009; 58:411–6.
11. Wiese L, Mejer N, Schönheyder HC, et al; Danish Staphylococcal Bacteraemia Study Group. A nationwide study of comorbidity and risk of reinfection after *Staphylococcus aureus* bacteraemia. *J Infect* 2013; 67:199–205.

12. Souli M, Ruffin F, Choi SH, et al. Changing characteristics of *Staphylococcus aureus* bacteremia: results from a 21-year, prospective, longitudinal study. *Clin Infect Dis* 2019; 69:1868–77.
13. Choi SH, Ruffin F, Park L, et al. 1060. Risk factors for recurrent *Staphylococcus aureus* bacteremia. *Open Forum Infect Dis* 2018; 5(suppl\_1):S317.
14. Chong YP, Moon SM, Bang KM, et al. Treatment duration for uncomplicated *Staphylococcus aureus* bacteremia to prevent relapse: analysis of a prospective observational cohort study. *Antimicrob Agents Chemother* 2013; 57:1150–6.
15. Friedman ND, Kaye KS, Stout JE, et al. Health care-associated bloodstream infections in adults: a reason to change the accepted definition of community-acquired infections. *Ann Intern Med* 2002; 137:791–7.
16. Sharma-Kuinkel BK, Rude TH, Fowler VG Jr. Pulse field gel electrophoresis. *Methods Mol Biol* 2016; 1373:117–30.
17. Tenover FC, Arbeit RD, Goering RV, et al. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995; 33:2233–9.
18. Hooton TM, Gupta K. Recurrent simple cystitis in women. In: Calderwood SB, ed. UpToDate. Waltham, MA: UpToDate Inc.
19. Johnson JR. Reinfection versus relapse in urinary tract infection. *Clin Infect Dis* 2005; 40: 495; author reply -6.
20. Mathema B, Mediavilla J, Kreiswirth BN. Sequence analysis of the variable number tandem repeat in *Staphylococcus aureus* protein A gene: spa typing. *Methods Mol Biol* 2008; 431:285–305.
21. Lindsay JA. Genomic variation and evolution of *Staphylococcus aureus*. *Int J Med Microbiol* 2010; 300:98–103.
22. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Series B* 1995; 57: 289–300.
23. Gualandi N, Mu Y, Bamberg WM, et al. Racial disparities in invasive methicillin-resistant *Staphylococcus aureus* infections, 2005–2014. *Clin Infect Dis* 2018; 67: 1175–81.
24. Klevens RM, Morrison MA, Nadle J, et al; Active Bacterial Core surveillance (ABCs) MRSA Investigators. Invasive methicillin-resistant *Staphylococcus aureus* infections in the United States. *JAMA* 2007; 298:1763–71.
25. See I, Wesson P, Gualandi N, et al. Socioeconomic factors explain racial disparities in invasive community-associated methicillin-resistant *Staphylococcus aureus* disease rates. *Clin Infect Dis* 2017; 64:597–604.
26. Bagger JP, Zindrou D, Taylor KM. Postoperative infection with methicillin-resistant *Staphylococcus aureus* and socioeconomic background. *Lancet* 2004; 363:706–8.
27. Tosas Auguet O, Betley JR, Stabler RA, et al. Evidence for community transmission of community-associated but not health-care-associated methicillin-resistant *Staphylococcus Aureus* strains linked to social and material deprivation: spatial analysis of cross-sectional data. *PLoS Med* 2016; 13:e1001944.
28. Gorwitz RJ, Kruszon-Moran D, McAllister SK, et al. Changes in the prevalence of nasal colonization with *Staphylococcus aureus* in the United States, 2001–2004. *J Infect Dis* 2008; 197: 1226–34.
29. Neves FPG, Marlow MA, Rezende-Pereira G, et al. Differences in gram-positive bacterial colonization and antimicrobial resistance among children in a high income inequality setting. *BMC Infect Dis* 2019; 19:478.
30. Freitas EA, Harris RM, Blake RK, Salgado CD. Prevalence of USA300 strain type of methicillin-resistant *Staphylococcus aureus* among patients with nasal colonization identified with active surveillance. *Infect Control Hosp Epidemiol* 2010; 31:469–75.
31. Cyr DD, Allen AS, Du GJ, et al. Evaluating genetic susceptibility to *Staphylococcus aureus* bacteremia in African Americans using admixture mapping. *Genes Immun* 2017; 18:95–9.
32. DeLorenze GN, Nelson CL, Scott WK, et al. Polymorphisms in HLA class II genes are associated with susceptibility to *Staphylococcus aureus* infection in a white population. *J Infect Dis* 2016; 213:816–23.
33. Albertus P, Morgenstern H, Robinson B, Saran R. Risk of ESRD in the United States. *Am J Kidney Dis* 2016; 68:862–72.
34. Tomson CR, Foley RN, Li Q, Gilbertson DT, Xue JL, Collins AJ. Race and end-stage renal disease in the United States Medicare population: the disparity persists. *Nephrology (Carlton)* 2008; 13:651–6.
35. Saunders MR, Lee H, Maene C, Schuble T, Cagney KA. Proximity does not equal access: racial disparities in access to high quality dialysis facilities. *J Racial Ethn Health Disparities* 2014; 1:291–9.
36. Corl K, Levy M, Phillips G, Terry K, Friedrich M, Trivedi AN. Racial and ethnic disparities in care following the New York State sepsis initiative. *Health Aff (Millwood)* 2019; 38:1119–26.
37. Marzec NS, Bessesen MT. Risk and outcomes of methicillin-resistant *Staphylococcus aureus* (MRSA) bacteremia among patients admitted with and without MRSA nares colonization. *Am J Infect Control* 2016; 44:405–8.



38. Wertheim HF, Vos MC, Ott A, et al. Risk and outcome of nosocomial *Staphylococcus aureus* bacteraemia in nasal carriers versus non-carriers. *Lancet* **2004**; 364:703–5.
39. von Eiff C, Becker K, Machka K, Stammer H, Peters G. Nasal carriage as a source of *Staphylococcus aureus* bacteremia. Study Group. *N Engl J Med* **2001**; 344:11–6.
40. Appay V, Rowland-Jones SL. RANTES: a versatile and controversial chemokine. *Trends Immunol* **2001**; 22:83–7.
41. McNicholas S, Talento AF, O’Gorman J, et al. Cytokine responses to *Staphylococcus aureus* bloodstream infection differ between patient cohorts that have different clinical courses of infection. *BMC Infect Dis* **2014**; 14:580.